

Vitamin C and Immune Function

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Introduction

This non-data review looks at vitamin C's physiological role in immune cells and immunity, as a whole.

Conclusions

According to this non-data review, vitamin C is used in large quantities by immune cells in both main branches of the immune system (innate or adaptive). Vitamin C is used to neutralize high levels of reactive oxygens species when immune cells phagocytose (consume and destroy) pathogens to protect the cell from causing damage to itself - as well as preserving the apoptotic (programmed cell death) pathway for proper clearance of damaged immune cells. In wound healing, it allows more stable collagen to be produced and put down in the extracellular matrix. Not only that, it balances cell to cell communication for inflammation, as well as allows immune cells to migrate to their destinations appropriately.

Amendments

This review is partly written by someone working in the pharmaceutical industry.



nutrients



Review

Vitamin C and Immune Function

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Abstract: Vitamin C is an essential micronutrient for humans, with pleiotropic functions related to its ability to donate electrons. It is a potent antioxidant and a cofactor for a family of biosynthetic and gene regulatory enzymes. Vitamin C contributes to immune defense by supporting various cellular functions of both the innate and adaptive immune system. Vitamin C supports epithelial barrier function against pathogens and promotes the oxidant scavenging activity of the skin, thereby potentially protecting against environmental oxidative stress. Vitamin C accumulates in phagocytic cells, such as neutrophils, and can enhance chemotaxis, phagocytosis, generation of reactive oxygen species, and ultimately microbial killing. It is also needed for apoptosis and clearance of the spent neutrophils from sites of infection by macrophages, thereby decreasing necrosis/NETosis and potential tissue damage. The role of vitamin C in lymphocytes is less clear, but it has been shown to enhance differentiation and proliferation of B- and T-cells, likely due to its gene regulating effects. Vitamin C deficiency results in impaired immunity and higher susceptibility to infections. In turn, infections significantly impact on vitamin C levels due to enhanced inflammation and metabolic requirements. Furthermore, supplementation with vitamin C appears to be able to both prevent and treat respiratory and systemic infections. Prophylactic prevention of infection requires dietary vitamin C intakes that provide at least adequate, if not saturating plasma levels (i.e., 100–200 mg/day), which optimize cell and tissue levels. In contrast, treatment of established infections requires significantly higher (gram) doses of the vitamin to compensate for the increased inflammatory response and metabolic demand.

Keywords: ascorbate; ascorbic acid; immunity; immune system; neutrophil function; microbial killing; lymphocytes; infection; vitamin C

1. Introduction

The immune system is a multifaceted and sophisticated network of specialized organs, tissues, cells, proteins, and chemicals, which has evolved in order to protect the host from a range of pathogens, such as bacteria, viruses, fungi, and parasites, as well as cancer cells [1]. It can be divided into epithelial barriers, and cellular and humoral constituents of either innate (non-specific) and acquired (specific) immunity [1]. These constituents interact in multiple and highly complex ways. More than half a century of research has shown vitamin C to be a crucial player in various aspects of the immune system, particularly immune cell function [2,3].

Vitamin C is an essential nutrient which cannot be synthesized by humans due to loss of a key enzyme in the biosynthetic pathway [4,5]. Severe vitamin C deficiency results in the potentially fatal disease scurvy [6]. Scurvy is characterized by weakening of collagenous structures, resulting in poor wound healing, and impaired immunity. Individuals with scurvy are highly susceptible to potentially fatal infections such as pneumonia [7]. In turn, infections can significantly impact on vitamin C levels due to enhanced inflammation and metabolic requirements. Early on, it was noted that scurvy often

followed infectious epidemics in populations [7], and cases of scurvy have been reported following respiratory infection [8]. This is particularly apparent for individuals who are already malnourished.

Although the amount of vitamin C required to prevent scurvy is relatively low (i.e., ~10 mg/day) [9], the recommended dietary intakes for vitamin C are up to one hundred-fold higher than that for many other vitamins [10]. A diet that supplies 100–200 mg/day of vitamin C provides adequate to saturating plasma concentrations in healthy individuals and should cover general requirements for the reduction of chronic disease risk [11,12]. Due to the low storage capacity of the body for the water-soluble vitamin, a regular and adequate intake is required to prevent hypovitaminosis C. Epidemiological studies have indicated that hypovitaminosis C (plasma vitamin C < 23 µmol/L) is relatively common in Western populations, and vitamin C deficiency (<11 µmol/L) is the fourth leading nutrient deficiency in the United States [13,14]. There are several reasons why vitamin C dietary recommendations are not met, even in countries where food availability and supply would be expected to be sufficient. These include poor dietary habits, life-stages and/or lifestyles either limiting intakes or increasing micronutrient requirements (e.g., smoking and alcohol or drug abuse), various diseases, exposure to pollutants and smoke (both active and passive), and economic reasons (poor socioeconomic status and limited access to nutritious food) [15,16]. Even otherwise 'healthy' individuals in industrialized countries can be at risk due to lifestyle-related factors, such as those on a diet or eating an unbalanced diet, and people facing periods of excessive physical or psychological stress [15,16].

Vitamin C has a number of activities that could conceivably contribute to its immune-modulating effects. It is a highly effective antioxidant, due to its ability to readily donate electrons, thus protecting important biomolecules (proteins, lipids, carbohydrates, and nucleic acids) from damage by oxidants generated during normal cell metabolism and through exposure to toxins and pollutants (e.g., cigarette smoke) [17]. Vitamin C is also a cofactor for a family of biosynthetic and gene regulatory monooxygenase and dioxygenase enzymes [18,19]. The vitamin has long been known as a cofactor for the lysyl and prolyl hydroxylases required for stabilization of the tertiary structure of collagen, and is a cofactor for the two hydroxylases involved in carnitine biosynthesis, a molecule required for transport of fatty acids into mitochondria for generation of metabolic energy (Figure 1) [19].

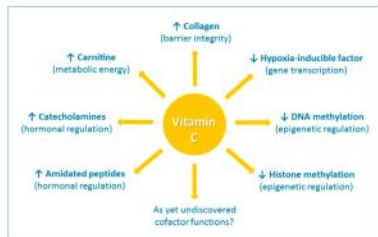


Figure 1. The enzyme cofactor activities of vitamin C. Vitamin C is a cofactor of a family of biosynthetic and gene regulatory monooxygenase and dioxygenase enzymes. These enzymes are involved in the synthesis of collagen, carnitine, catecholamine hormones, e.g., norepinephrine, and amidated peptide hormones, e.g., vasopressin. These enzymes also hydroxylate transcription factors, e.g., hypoxia-inducible factor 1 α , and methylated DNA and histones, thus playing a role in gene transcription and epigenetic regulation. ↑ indicates an increase and ↓ indicates a decrease.

Vitamin C is also a cofactor for the hydroxylase enzymes involved in the synthesis of catecholamine hormones, e.g., norepinephrine, and amidated peptide hormones e.g., vasopressin, which are central to the cardiovascular response to severe infection [20]. Furthermore, research over the past 15 years or so has uncovered new roles for vitamin C in the regulation of gene transcription and cell signaling pathways through regulation of transcription factor activity and epigenetic marks (Figure 1) [21,22]. For example, the asparagyl and prolyl hydroxylases required for the downregulation of the pleiotropic transcription factor hypoxia-inducible factor-1 α (HIF-1 α) utilize vitamin C as a cofactor [21]. Recent research has also indicated an important role for vitamin C in regulation of DNA and histone methylation by acting as a cofactor for enzymes which hydroxylate these epigenetic marks [22].

Our review explores the various roles of vitamin C in the immune system, including barrier integrity and leukocyte function, and discusses potential mechanisms of action. We discuss the relevance of the immune-modulating effects of vitamin C in the context of infections and conditions leading to vitamin C insufficiency.

2. Barrier Integrity and Wound Healing

The skin has numerous essential functions, the primary of which is to act as a barrier against external insults, including pathogens. The epidermal layer is highly cellular, comprising primarily keratinocytes, whilst the dermal layer comprises fibroblasts which secrete collagen fibers, the major component of the dermis [23]. Skin contains millimolar concentrations of vitamin C, with higher levels found in the epidermis than the dermis [24–26]. Vitamin C is actively accumulated into the epidermal and dermal cells via the two sodium-dependent vitamin C transporter (SVCT) isoforms 1 and 2 [27], suggesting that the vitamin has crucial functions within the skin. Clues to the role of vitamin C in the skin come from the symptoms of the vitamin C deficiency disease scurvy, which is characterized by bleeding gums, bruising, and impaired wound healing [28,29]. These symptoms are thought to be a result of the role of vitamin C as a co-factor for the prolyl and lysyl hydroxylase enzymes that stabilize the tertiary structure of collagen (Table 1) [30]. Further research has shown that vitamin C can also increase collagen gene expression in fibroblasts [31–35].

Table 1. Role of vitamin C in immune defense.

Immune System	Function of Vitamin C	Refs.
Epithelial barriers	Enhances collagen synthesis and stabilization	[30–35]
	Protects against ROS-induced damage ¹	[36–40]
	Enhances keratinocyte differentiation and lipid synthesis	[41–45]
	Enhances fibroblast proliferation and migration	[46,47]
Phagocytes (neutrophils, macrophages)	Shortens time to wound healing in patients	[48,49]
	Acts as an antioxidant/electron donor	[50–53]
	Enhances motility/chemotaxis	[54–63]
	Enhances phagocytosis and ROS generation	[64–71]
	Enhances microbial killing	[54,55,57,58,70,72]
B- and T-lymphocytes	Facilitates apoptosis and clearance	[71,73,74]
	Decreases necrosis/NEtosis	[75,76]
Inflammatory mediators	Enhances differentiation and proliferation	[62,65,76–82]
	Enhances antibody levels	[78,83–85]
Inflammatory mediators	Modulates cytokine production	[75,77,86–88]
	Decreases histamine levels	[86,89–101]

¹ ROS, reactive oxygen species; NET, neutrophil extracellular trap. Note that many of these studies comprised marginal or deficient vitamin C status at baseline. Supplementation in situations of adequate vitamin C status may not have comparable effects.

Vitamin C intervention studies in humans (using both dietary and gram doses of vitamin C) have shown enhanced vitamin C uptake into skin cells [26,36] and enhanced oxidant scavenging activity

Barrier Integrity and Wound Healing

Skin is made up three primary sections, but only two are discussed here: the dermis and epidermis. The epidermis is the section we can see with our eye when we look at our skin - it is made up many skin cells (keratinocytes). The dermis (or dermal layer) is just below the epidermis and consists of non-skin fibroblasts (cells that produce collagen). More Vitamin C is located in the epidermis (likely, because there are more skin cells there).

Vitamin C finds its way into the skin cells by a sodium dependent vitamin C transporter (SVCT) that allows ascorbic acid (vitamin C) into the cell when accompanied by two sodium ions.

A lack of vitamin C leads to impaired wound healing (like that seen in Scurvy).

Vitamin C (ascorbic acid) is used in an enzyme reaction created by two enzymes - prolyl and lysyl hydroxylase - that hydroxylate the amino acids making up the triple helical structure of collagen. In layman terms, the collagen protein has a chemical alteration made to it (hydroxylation) that makes it more stable and less likely to be degraded.

Also, vitamin C stimulates collagen gene expression in fibroblasts (in the dermis).

Vitamin C can also play a role as an antioxidant ("calming" reactive/destructive molecules within the cell).

Vitamin C increases lipid synthesis for the keratinocytes and these lipids are packed between the cells to create a barrier (called the lamellae) - the denser the barrier, the better - so, more lipids/fats organized in that location, the better. These cells, with vitamin C, also increase the number of tight junction proteins between them. Tight junction proteins are proteins that anchor one cell to the next, closing the space between cells.

There is evidence that vitamin C does not play a role in normal collagen homeostasis (creating and degrading collagen), except when damage occurs to the skin, in which case vitamin C becomes important as collagen synthesis is impeded (through its stabilizing ability in collagen, discussed earlier). Vitamin C, in wound healing, also decreases pro-inflammatory signaling and enhances wound healing. It also encourages fibroblast (collagen producing cells) to multiply and migrate to the wound area.

Larger amounts of vitamin C (500 mg/day, for example) are associated with faster wound healing.

Vitamin C also plays a role in neutrophil (innate immune cell) migration to the wounded area and disinfecting it - this is discussed in the next section.

of the skin [36,37]. The elevated antioxidant status of the skin following vitamin C supplementation could potentially protect against oxidative stress induced by environmental pollutants [38,39]. The antioxidant effects of vitamin C are likely to be enhanced in combination with vitamin E [40,102].

Cell culture and preclinical studies have indicated that vitamin C can enhance epithelial barrier functions via a number of different mechanisms. Vitamin C supplementation of keratinocytes in culture enhances differentiation and barrier function via modulating signaling and biosynthetic pathways, with resultant elevations in barrier lipid synthesis [41–45]. Dysfunctional epithelial barrier function in the lungs of animals with serious infection can be restored by administration of vitamin C [74]. This was attributed to enhanced expression of tight junction proteins and prevention of cytoskeletal rearrangements.

Animal studies using the vitamin C-dependent Gulo knockout mouse indicated that deficiency did not affect the formation of collagen in the skin of unchallenged mice [103]; however, following full thickness excisional wounding there was significantly decreased collagen formation in vitamin C-deficient mice [46]. This finding is in agreement with an earlier study carried out with scurbutic guinea pigs [104]. Thus, vitamin C appears to be particularly essential during wound healing, also decreasing the expression of pro-inflammatory mediators and enhancing the expression of various wound healing mediators [46]. Fibroblast cell culture experiments have also indicated that vitamin C can alter gene expression profiles within dermal fibroblasts, promoting fibroblast proliferation and migration which is essential for tissue remodeling and wound healing [46,47]. Following surgery, patients require relatively high intakes of vitamin C in order to normalize their plasma vitamin C status (e.g., ≥ 500 mg/day) [105], and administration of antioxidant micronutrients, including vitamin C, to patients with disorders in wound healing can shorten the time to wound closure [48,49,106,107].

Leukocytes, particularly neutrophils and monocyte-derived macrophages, are major players in wound healing [108]. During the initial inflammatory stage, neutrophils migrate to the wound site in order to sterilize it via the release of reactive oxygen species (ROS) and antimicrobial proteins [109]. The neutrophils eventually undergo apoptosis and are cleared by macrophages, resulting in resolution of the inflammatory response. However, in chronic, non-healing wounds, such as those observed in diabetics, the neutrophils persist and instead undergo necrotic cell death which can perpetuate the inflammatory response and hinder wound healing [109,110]. Vitamin C is thought to influence several important aspects of neutrophil function: migration in response to inflammatory mediators (chemotaxis), phagocytosis and killing of microbes, and apoptosis and clearance by macrophages (see below).

3. Vitamin C and Leukocyte Function

Leukocytes, such as neutrophils and monocytes, actively accumulate vitamin C against a concentration gradient, resulting in values that are 50- to 100-fold higher than plasma concentrations [111–113]. These cells accumulate maximal vitamin C concentrations at dietary intakes of ~ 100 mg/day [114,115], although other body tissues likely require higher intakes for saturation [116,117]. Neutrophils accumulate vitamin C via SVCT2 and typically contain intracellular levels of at least 1 mM [111,118]. Following stimulation of their oxidative burst neutrophils can further increase their intracellular concentration of vitamin C through the non-specific uptake of the oxidized form, dehydroascorbate (DHA), via glucose transporters (GLUT) [118,119]. DHA is then rapidly reduced to ascorbate intracellularly, to give levels of about 10 mM [119]. It is believed that the accumulation of such high vitamin C concentrations indicates important functions within these cells.

Accumulation of millimolar concentrations of vitamin C into neutrophils, particularly following activation of their oxidative burst, is thought to protect these cells from oxidative damage [119].

Vitamin C and Leukocyte Function

Leukocytes are immune cells. Vitamin C is taken up by these cells at a mass concentration up to 100 times higher than that found in the blood. They take up vitamin C through the

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Accumulation of millimolar concentrations of vitamin C into neutrophils, particularly following activation of their oxidative burst, is thought to protect these cells from oxidative damage [119]. Vitamin C is a potent water-soluble antioxidant that can scavenge numerous reactive oxidants and can also regenerate the important cellular and membrane antioxidants glutathione and vitamin E [120]. Upon phagocytosis or activation with soluble stimulants, vitamin C is depleted from neutrophils in an oxidant-dependent manner [50–53]. An alteration in the balance between oxidant generation and

Vitamin C and Leukocyte Function

Leukocytes are immune cells. Vitamin C is taken up by these cells at a mass concentration up to 100 times higher than that found in the blood. They take up vitamin C through the aforementioned SVCT (vitamin C transporter). When neutrophils (a type of leukocyte) initiate an "oxidative burst" wherein they bombard a pathogen (ex. bacteria) they have enveloped with oxidative stress (molecules that destroy other molecules). However, since these molecules are so reactive, it is believed that vitamin C concentrations neutralize this effect in the cell to protect the cell, itself. As such, during these oxidative bursts, another form of vitamin C is taken up by GLUT receptors to bolster vitamin C levels in the cell.

Running out of vitamin C in the cells leads to a greater "pro-inflammatory" reaction by the cell, causing the cells to release more inflammatory signals and produce more oxidative molecules. Oxidative molecules within the cell can also lead to damage of the motility (mobility) parts of the cell (like the cell membrane made of lipids, which should be intact to allow fluidity in movement).

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antioxidant defenses can lead to alterations in multiple signaling pathways, with the pro-inflammatory transcription factor nuclear factor κ B (NF κ B) playing a central role [121]. Oxidants can activate NF κ B, which triggers a signaling cascade leading to continued synthesis of oxidative species and other inflammatory mediators [122,123]. Vitamin C has been shown to attenuate both oxidant generation and NF κ B activation in dendritic cells in vitro, and NF κ B activation in neutrophils isolated from septic Gulo knockout mice [75,124]. Thiol-containing proteins can be particularly sensitive to redox alterations within cells and are often central to the regulation of redox-related cell signaling pathways [125]. Vitamin C-dependent modulation of thiol-dependent cell signaling and gene expression pathways has been reported in T-cells [126,127].

Thus, vitamin C could modulate immune function through modulation of redox-sensitive cell signaling pathways or by directly protecting important cell structural components. For example, exposure of neutrophils to oxidants can inhibit motility of the cells, which is thought to be due to oxidation of membrane lipids and resultant effects on cell membrane fluidity [63]. Neutrophils contain high levels of polyunsaturated fatty acids in their plasma membranes, and thus improvements in neutrophil motility observed following vitamin C administration (see below) could conceivably be attributed to oxidant scavenging as well as regeneration of vitamin E [120].

3.1. Neutrophil Chemotaxis

Neutrophil infiltration into infected tissues is an early step in innate immunity. In response to pathogen- or host-derived inflammatory signals (e.g., *N*-formylmethionyl-leucyl-phenylalanine (fMLP), interleukin (IL)-8, leukotriene B₄, and complement component C5a), marginated neutrophils literally swarm to the site of infection [128]. Migration of neutrophils in response to chemical stimuli is termed chemotaxis, while random migration is termed chemokinesis (Figure 2). Neutrophils express more than 30 different chemokine and chemoattractant receptors in order to sense and rapidly respond to tissue damage signals [128]. Early studies carried out in scorbutic guinea pigs indicated impaired leukocyte chemotactic response compared with leukocytes isolated from guinea pigs supplemented with adequate vitamin C in their diet (Table 1) [54–56,64]. These findings suggest that vitamin C deficiency may impact on the ability of phagocytes to migrate to sites of infection.

Patients with severe infection exhibit compromised neutrophil chemotactic ability [129–132]. This neutrophil "paralysis" is believed to be partly due to enhanced levels of anti-inflammatory and immune-suppressive mediators (e.g., IL-4 and IL-10) during the compensatory anti-inflammatory response observed following initial hyper-stimulation of the immune system [133]. However, it is also possible that vitamin C depletion, which is prevalent during severe infection [20], may contribute. Studies in the 1980s and 1990s indicated that patients with recurrent infections had impaired leukocyte chemotaxis, which could be restored in response to supplementation with gram doses of vitamin C [57–60,65–67]. Furthermore, supplementation of neonates with suspected sepsis with 400 mg/day vitamin C dramatically improved neutrophil chemotaxis [134].

Recurrent infections can also result from genetic disorders of neutrophil function, such as chronic granulomatous disease (CGD), an immunodeficiency disease resulting in defective leukocyte generation of ROS [135], and Chediak-Higashi syndrome (CHS), a rare autosomal recessive disorder affecting vesicle trafficking [136]. Although vitamin C administration would not be expected to affect the underlying defects of these genetic disorders, it may support the function of redundant antimicrobial mechanisms in these cells. For example, patients with CGD showed improved leukocyte chemotaxis following supplementation with gram doses of vitamin C administered either enterally or parenterally [137–139]. This was associated with decreased infections and clinical improvement [137,138]. A mouse model of CHS showed improved neutrophil chemotaxis following vitamin C supplementation [140], and neutrophils isolated from two children with CHS showed improved chemotaxis following supplementation with 200–500 mg/day vitamin C [141,142], although this effect has not been observed in all cases [140,143]. The vitamin C-dependent enhancement of

Neutrophil Chemotaxis

Neutrophils are leukocytes/immune cells. Chemotaxis is the response by immune cells to a signal (chemokine) released by other cells and the migration of those immune cells toward the signal. Neutrophils swarm to areas of damage or infection. Vitamin C deficiency is linked to decreased ability for neutrophils to migrate/move to sites of infection.

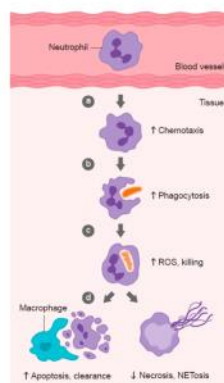
Neutrophil "paralysis" (lack of movement to infection site) is thought to be due to heightened anti-inflammatory signaling (chemokines) in response to an initial hyper-inflammatory environment (also mediated by chemokines, but they are pro-inflammatory).

There is some evidence that vitamin C rejuvenates chemotaxis. Vitamin C is thought to improve microtubule assembly (this is the structural units of the cell) by improving their stability in the cell and the ability to transport and move things within the cell.

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chemotaxis was thought to be mediated in part via effects on microtubule assembly [144,145], and more recent research has indicated that intracellular vitamin C can stabilize microtubules [146].



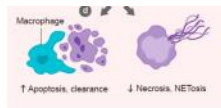


Figure 2. Role of vitamin C in phagocyte function. Vitamin C has been shown to: (a) enhance neutrophil migration in response to chemotactants (chemotaxis), (b) enhance engulfment (phagocytosis) of microbes, and (c) stimulate reactive oxygen species (ROS) generation and killing of microbes. (d) Vitamin C supports caspase-dependent apoptosis, enhancing uptake and clearance by macrophages, and inhibits necrosis, including NETosis, thus supporting resolution of the inflammatory response and attenuating tissue damage.

Supplementation of healthy volunteers with dietary or gram doses of vitamin C has also been shown to enhance neutrophil chemotactic ability [61–63,147]. Johnston et al., proposed that the antihistamine effect of vitamin C correlated with enhanced chemotaxis [61]. In participants who had inadequate vitamin C status (i.e., <50 µM), supplementation with a dietary source of vitamin C (providing ~250 mg/day) resulted in a 20% increase in neutrophil chemotaxis [147]. Furthermore, supplementation of elderly women with 1 g/day vitamin C, in combination with vitamin E, enhanced neutrophil functions, including chemotaxis [148]. Thus, members of the general population may benefit from improved immune cell function through enhanced vitamin C intake, particularly if they have inadequate vitamin C status, which can be more prevalent in the elderly. However, it should be noted that it is not yet certain to what extent improved ex vivo leukocyte chemotaxis translates into improved in vivo immune function.

3.2. Phagocytosis and Microbial Killing

Once neutrophils have migrated to the site of infection, they proceed to engulf the invading pathogens (Figure 2). Various intracellular granules are mobilized and fuse with the phagosome,

emptying their arsenal of antimicrobial peptides and proteins into the phagosome [149]. Components of the nicotinamide adenine dinucleotide phosphate (NADPH) oxidase assemble in the phagosomal membrane and generate superoxide, the first in a long line of ROS generated by neutrophils to kill pathogens. The enzyme superoxide dismutase converts superoxide to hydrogen peroxide, which can then be utilized to form the oxidant hypochlorous acid via the azurophilic granule enzyme myeloperoxidase [149]. Hypochlorous acid can further react with amines to form secondary oxidants known as chloramines. These various neutrophil-derived oxidants have different reactivities and specificities for biological targets, with protein thiol groups being particularly susceptible.

Neutrophils isolated from scorbutic guinea pigs exhibit a severely impaired ability to kill microbes [54,55,70], and studies have indicated impaired phagocytosis and/or ROS generation in neutrophils from scorbutic compared with ascorbate replete animals [68–70]. Generation of ROS by neutrophils from volunteers with inadequate vitamin C status can be enhanced by 20% following supplementation with a dietary source of vitamin C [147], and increases in both phagocytosis and oxidant generation were observed following supplementation of elderly participants with a combination of vitamins C and E [148]. Patients with recurrent infections [57,58,66,67,72], or the genetic conditions CGD or CHS [138,139,141,143,150], have impaired neutrophil bacterial killing and/or phagocytosis, which can be significantly improved following supplementation with gram doses of vitamin C, resulting in long-lasting clinical improvement. A couple of studies, however, showed no improvement of ex vivo anti-fungal or anti-bacterial activity in neutrophils isolated from CGD or CHS patients supplemented with vitamin C [140,151]. The reason for these differences is not clear, although it may depend on the baseline vitamin C level of the patients, which is not assessed in most cases. Furthermore, different microbes have variable susceptibility to the oxidative and non-oxidative anti-microbial mechanisms of neutrophils. For example, *Staphylococcus aureus* is susceptible to oxidative mechanisms, whereas other microorganisms are more susceptible to non-oxidative mechanisms [152]. Therefore, the type of microbe used to assess the ex vivo neutrophil functions could influence the findings.

Patients with severe infection (sepsis) exhibit a decreased ability to phagocytose microbes and a diminished ability to generate ROS [153]. Decreased neutrophil phagocytosis was associated with enhanced patient mortality [154]. Interestingly, Stephan et al. [155] observed impaired neutrophil killing activity in critically ill patients prior to acquiring nosocomial infections, suggesting that critical illness itself, without prior infection, can also impair neutrophil function. This resulted in subsequent susceptibility to hospital-acquired infections. Impaired phagocytic and oxidant-generating capacity of leukocytes in patients with severe infection has been attributed to the compensatory anti-inflammatory response, resulting in enhanced levels of immunosuppressive mediators such as IL-10 [133], as well as to the hypoxic conditions of inflammatory sites, which diminishes substrate for ROS generation [156]. Another explanation is the larger numbers of immature neutrophils released from the bone marrow due to increased demands during severe infection. These immature 'band' cells have decreased functionality compared with differentiated neutrophils [157]. Thus, conflicting findings in severe infection could be due to variability in the total numbers of underactive immature neutrophils compared with activated fully-differentiated neutrophils [158,159]. Despite displaying an activated basal state, the mature neutrophils from patients with severe infection do not generate ROS to the same extent as healthy neutrophils following ex vivo stimulation [160]. The effect of vitamin C supplementation on phagocytosis, oxidant generation, and microbial killing by leukocytes from septic patients has not yet been explored.

3.3. Neutrophil Apoptosis and Clearance

Following microbial phagocytosis and killing, neutrophils undergo a process of programmed cell death called apoptosis [161]. This process facilitates subsequent phagocytosis and clearance of the spent neutrophils from sites of inflammation by macrophages, thus supporting resolution of inflammation and preventing excessive tissue damage (Figure 2). Caspases are key effector enzymes

in the apoptotic process, culminating in phosphatidyl serine exposure, thus marking the cells for uptake and clearance by macrophages [162]. Interestingly, caspases are thiol-dependent enzymes, making them very sensitive to inactivation by ROS generated by activated neutrophils [163,164]. Thus, vitamin C may be expected to protect the oxidant-sensitive caspase-dependent apoptotic process following activation of neutrophils. In support of this premise, in vitro studies have shown that loading human neutrophils with vitamin C can enhance *Escherichia coli*-mediated apoptosis of the neutrophils (Table 1) [71]. Peritoneal neutrophils isolated from vitamin C-deficient Gulo mice exhibited attenuated apoptosis [75], and instead underwent necrotic cell death [73]. These vitamin C-deficient neutrophils

Phagocytosis and Microbial Killing

Neutrophils (immune cells) can undergo the process of phagocytosis (consuming pathogens - bacteria) and in doing so, fuse vesicles of acidic components and oxidation molecules with a vesicle containing the pathogen. This allows the pathogen to be destroyed by these toxic systems.

Vitamin C deficient guinea pigs show an impaired ability to phagocytose (consume pathogens) and to produce reactive oxygen species (oxidizing molecules). This effect may only be seen with particular microbes/pathogens that are susceptible to reactive oxygen specie mediated destruction).

Neutrophil Apoptosis and Clearance

Apoptosis is programmed cell death. Once neutrophils destroy pathogens, they can self induce an apoptotic signal that also recruits other immune cells to consume the remaining debris, thereby creating a clean context for death. This self induced apoptosis is mediated by caspases, which are proteins that, once activated, send a cascade of signals throughout the cell and lead to cell destruction by destroying the mitochondria and nucleus of the cell.

Apoptosis is beneficial, because the internal parts of the cell can lead to a heightened, unwarranted immune response if it is exposed to the extracellular area/fluid (this is called necrosis).

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Numerous studies have reported attenuated neutrophil apoptosis in patients with severe infection compared with control participants [165–172]. The delayed apoptosis appears to be related to disease severity and is thought to be associated with enhanced tissue damage observed in patients with sepsis [173,174]. Immature ‘band’ neutrophils released during severe infection were also found to be resistant to apoptosis and had longer life spans [157]. Plasma from septic patients has been found to suppress apoptosis in healthy neutrophils, suggesting that pro-inflammatory cytokines were responsible for the increased in vivo survival of neutrophils during inflammatory conditions [165,174–176]. Interestingly, high-dose vitamin C administration has been shown to modulate cytokine levels in patients with cancer [177] and, although this has not yet been assessed in patients with severe infection, could conceivably be another mechanism by which vitamin C may modulate neutrophil function in these patients. To date, only one study has investigated the effect of vitamin C supplementation on neutrophil apoptosis in septic patients [178]. Intravenous supplementation of septic abdominal surgery patients with 450 mg/day vitamin C was found to decrease caspase-3 protein levels and, thus was presumed to have an anti-apoptotic effect on peripheral blood neutrophils. However, caspase activity and apoptosis of the neutrophils following activation was not assessed. Furthermore, circulating neutrophils may not reflect the activation status of neutrophils at inflammatory tissue loci. Clearly, more studies need to be undertaken to tease out the role of vitamin C in neutrophil apoptosis and clearance from inflammatory loci.

3.4. Neutrophil Necrosis and NETosis

Neutrophils that fail to undergo apoptosis instead undergo necrotic cell death (Figure 2). The subsequent release of toxic intracellular components, such as proteases, can cause extensive tissue damage [179,180]. One recently discovered form of neutrophil death has been termed NETosis. This results from the release of ‘neutrophil extracellular traps’ (NETs) comprising neutrophil DNA, histones, and enzymes [181]. Although NETs have been proposed to comprise a unique method of microbial killing [182,183], they have also been implicated in tissue damage and organ failure [184,185]. NET-associated histones can act as damage-associated molecular pattern proteins, activating the immune system and causing further damage [186]. Patients with sepsis, or who go on to develop sepsis, have significantly elevated levels of circulating cell-free DNA, which is thought to indicate NET formation [184,187].

Pre-clinical studies in vitamin C-deficient Gulo knockout mice indicated enhanced NETosis in the lungs of septic animals and increased circulating cell-free DNA [75]. The levels of these markers were attenuated in vitamin C sufficient animals or in deficient animals that were administered vitamin C (Table 1). The same investigators showed that in vitro supplementation of human neutrophils with vitamin C attenuated phorbol ester-induced NETosis [75]. Administration of gram doses of vitamin C to septic patients over four days, however, did not appear to decrease circulating cell-free DNA levels [188], although the duration of treatment may have been too short to see a sustained effect. It should be noted that cell-free DNA is not specific for neutrophil-derived DNA, as it may also derive from necrotic tissue; however, the association of neutrophil-specific proteins

which are proteins that, once activated, send a cascade of signals throughout the cell and lead to cell destruction by destroying the mitochondria and nucleus of the cell.

Apoptosis is beneficial, because the internal parts of the cell can lead to a heightened, unwarranted immune response if it is exposed to the extracellular area/fluid (this is called necrosis).

As Reactive Oxygen Species (ROS - oxidizing molecules) are generated for the use of destroying the pathogen (ex. bacteria), the oxidation sensitive caspases can be deactivated as their structure is compromised by ROS. As such, vitamin C can reduce the level of ROS, thereby protecting caspases for their proper pro-apoptotic effect. Delayed or impeded apoptosis is associated with disease severity, as well.

or enzymes, such as myeloperoxidase, with the DNA can potentially provide an indication of its source [184].

The transcription factor HIF-1 α facilitates neutrophil survival at hypoxic loci through delaying apoptosis [189]. Interestingly, vitamin C is a cofactor for the iron-containing dioxygenase enzymes that regulate the levels and activity of HIF-1 α [190]. These hydroxylase enzymes downregulate HIF-1 α activity by facilitating degradation of constitutively expressed HIF-1 α and decreasing binding of transcription coactivators. In vitamin C-deficient Gulo knockout mice, up-regulation of HIF-1 α was observed under normoxic conditions, along with attenuated neutrophil apoptosis and clearance by macrophages [73]. HIF-1 α has also been proposed as a regulator of NET generation by neutrophils [191], hence providing a potential mechanism by which vitamin C could downregulate NET generation by these cells [73].

3.5. Lymphocyte Function

Like phagocytes, B- and T-lymphocytes accumulate vitamin C to high levels via SVCT [192,193]. The role of vitamin C within these cells is less clear, although antioxidant protection has been suggested [194]. In vitro studies have indicated that incubation of vitamin C with lymphocytes promotes proliferation [76,77], resulting in enhanced antibody generation [78], and also provides resistance to various cell death stimuli [195]. Furthermore, vitamin C appears to have an important role in developmental differentiation and maturation of immature T-cells (Table 1) [76,79]. Similar proliferative and differentiation/maturation effects have been observed with mature and immature natural killer cells, respectively [196].

Early studies in guinea pigs showed enhanced mitotic activity of isolated peripheral blood lymphocytes following intraperitoneal vitamin C treatment, and enhanced humoral antibody levels during immunization [82–85]. Although one human intervention study has reported positive associations between antibody levels (immunoglobulin (Ig)M, (Ig)G, (Ig)A) and vitamin C supplementation [85], another has not [62]. Instead, Anderson and coworkers showed that oral and intravenous supplementation of low gram doses of vitamin C to children with asthma and healthy volunteers enhanced lymphocyte transformation, an ex vivo measure of mitogen-induced proliferation and enlargement of T-lymphocytes (Table 1) [62,63,81]. Administration of vitamin C to elderly people was also shown to enhance ex vivo lymphocyte proliferation [80], a finding confirmed using combinations of vitamin C with vitamins A and/or E [148,197]. Exposure to toxic chemicals can affect lymphocyte function, and both natural killer cell activity and lymphocyte blastogenic responses to T- and B-cell mitogens were restored to normal levels following vitamin C supplementation [198]. Although the human studies mentioned above are encouraging, it is apparent that more human intervention studies are needed to confirm these findings.

Recent research in wild-type and Gulo knockout mice indicated that parenteral administration of 200 mg/kg vitamin C modulated the immunosuppression of regulatory T-cells (Tregs) observed in sepsis [89]. Vitamin C administration enhanced Treg proliferation and inhibited the negative immunoregulation of Tregs by inhibiting the expression of specific transcription factors, antigens, and cytokines [89]. The mechanisms involved likely rely on the gene regulatory effects of vitamin C [79,89,199,200]. For example, recent research has implicated vitamin C in epigenetic regulation through its action as a cofactor for the iron-containing dioxygenases which hydroxylate methylated DNA and histones [22,201]. The ten-eleven translocation (TET) enzymes hydroxylate methylcytosine residues, which may act as epigenetic marks in their own right, and also facilitate removal of the methylated residues, an important process in epigenetic regulation [202]. Preliminary evidence indicates that vitamin C can regulate T-cell maturation via epigenetic mechanisms involving the TETs and histone demethylation [79,199,200]. It is likely that the cell signaling and gene regulatory functions of vitamin C, via regulation of transcription factors and epigenetic marks, play major roles in its immune-regulating functions.

Lymphocyte Function

Lymphocytes are a more advanced form of immune cell in that they are specialized and form antibodies, as well as specifically target a pathogen based on the pathogen's structure. These cells, much like innate immune cells (neutrophils, for example) suck up large amounts of vitamin C, as well. The reasons are more obscure, but it is believed it promotes their proliferation (multiplying into more numerous cells), as well as their ability to generate antibodies against specific pathogens. It is believed that vitamin C may play a role in epigenetic modifications (as a cofactor) to create these effects.

3.6. Inflammatory Mediators

Cytokines are important cell signaling molecules secreted by a variety of immune cells, both innate and adaptive, in response to infection and inflammation [1]. They comprise a broad range of molecules, including chemokines, interferons (IFNs), ILs, lymphokines, and TNFs, which modulate both humoral and cell-based immune responses, and regulate the maturation, growth, and responsiveness of specific cell populations. Cytokines can elicit pro-inflammatory or anti-inflammatory responses, and vitamin C appears to modulate systemic and leukocyte-derived cytokines in a complex manner.

Incubation of vitamin C with peripheral blood lymphocytes decreased lipopolysaccharide (LPS)-induced generation of the pro-inflammatory cytokines TNF- α and IFN- γ , and increased anti-inflammatory IL-10 production, while having no effect on IL-1 β levels [77]. Furthermore, in vitro addition of vitamin C to peripheral blood monocytes isolated from pneumonia patients decreased the generation of the pro-inflammatory cytokines TNF- α and IL-6 [86]. However, another study found that in vitro treatment of peripheral blood monocytes with vitamin C and/or vitamin E enhanced LPS-stimulated TNF- α generation, but did not affect IL-1 β generation [87]. Furthermore, incubation of vitamin C with virus-infected human and murine fibroblasts enhanced generation of antiviral IFN [91–93]. Supplementation of healthy human volunteers with 1 g/day vitamin C (with and without vitamin E) was shown to enhance peripheral blood mononuclear cell-derived IL-10, IL-1, and TNF- α following stimulation with LPS [87,94]. Thus, the effect of vitamin C on cytokine generation appears to depend on the cell type and/or the inflammatory stimulant. Recent research has indicated that vitamin C treatment of microglia, resident myeloid-derived macrophages in the central nervous system, attenuates activation of the cells and synthesis of the pro-inflammatory cytokines TNF, IL-6, and IL-1 β [90]. This is indicative of an anti-inflammatory phenotype.

Preclinical studies using Gulo knockout mice have highlighted the cytokine-modulating effects of vitamin C. Vitamin C-deficient Gulo knockout mice infected with influenza virus showed enhanced synthesis of the pro-inflammatory cytokines TNF- α and IL-1 α/β in their lungs, and decreased production of the anti-viral cytokine IFN- α/β [88]. Administration of vitamin C to Gulo mice with polynicrobial peritonitis resulted in decreased synthesis of the pro-inflammatory cytokines TNF- α and IL-1 β by isolated neutrophils [75]. Another study in septic Gulo mice administered 200 mg/kg parenteral vitamin C has shown decreased secretion of the inhibitory cytokines TGF- β and IL-10 by Tregs [89]. In this study, attenuated IL-4 secretion and augmented IFN- γ secretion was also observed, suggesting immune-modulating effects of vitamin C in sepsis. Overall, vitamin C appears to normalize cytokine generation, likely through its gene-regulating effects.

Histamine is an immune mediator produced by basophils, eosinophils, and mast cells during the immune response to pathogens and stress. Histamine stimulates vasodilation and increased capillary permeability, resulting in the classic allergic symptoms of runny nose and eyes. Studies using guinea pigs, a vitamin C-requiring animal model, have indicated that vitamin C depletion is associated with enhanced circulating histamine levels, and that supplementation of the animals with vitamin C resulted in decreased histamine levels [56,95–98]. Enhanced histamine generation was found to increase the utilization of vitamin C in these animals [96]. Consistent with the animal studies, human intervention studies with oral vitamin C (125 mg/day to 2 g/day) and intravenous vitamin C (7.5 g infusion) have reported decreased histamine levels [61,99–101], which was more apparent in patients with allergic compared with infectious diseases [101]. Although vitamin C has been proposed to ‘detoxify’ histamine [96,97], the precise mechanisms responsible for the in vivo decrease in histamine levels following vitamin C administration are currently unknown. Furthermore, effects of vitamin C supplementation on histamine levels are not observed in all studies [203].

4. Vitamin C Insufficiency Conditions

Numerous environmental and health conditions can have an impact on vitamin C status. In this section we discuss examples which also have a link with impaired immunity and increased susceptibility to infection. For example, exposure to air pollution containing oxidants, such as ozone

Inflammatory Mediators

Cytokines are communication proteins used between cells. Some are used to accentuate a pro-inflammatory signal and others are anti-inflammatory (meaning, inflammation recruits more inflammatory cells to an area of the body). There are conflicting studies, but no doubt vitamin C does change the way cells express cytokines (more or less inflammatory cytokines).

and nitrogen dioxide, can upset the oxidant-antioxidant balance within the body and cause oxidative stress [204]. Oxidative stress can also occur if antioxidant defenses are impaired, which may be the case when vitamin C levels are insufficient [205]. Air pollution can damage respiratory tract lining fluid and increase the risk of respiratory disease, particularly in children and the elderly [204,206] who are at risk of both impaired immunity and vitamin C insufficiency [14,204]. Vitamin C is a free-radical scavenger that can scavenge superoxide and peroxyl radicals, hydrogen peroxide, hypochlorous acid, and oxidant air pollutants [207,208]. The antioxidant properties of vitamin C enable it to protect lung cells exposed to oxidants and oxidant-mediated damage caused by various pollutants, heavy metals, pesticides, and xenobiotics [204,209].

Tobacco smoke is an underestimated pollutant in many parts of the world. Both smokers and passive smokers have lower plasma and leukocyte vitamin C levels than non-smokers [10,210,211], partly due to increased oxidative stress and to both a lower intake and a higher metabolic turnover of vitamin C compared to non-smokers [10,211–213]. Mean serum concentrations of vitamin C in adults who smoke have been found to be one-third lower than those of non-smokers, and it has been recommended that smokers should consume an additional 35 mg/day of vitamin C to ensure there is sufficient ascorbic acid to repair oxidant damage [10,14]. Vitamin C levels are also lower in children and adolescents exposed to environmental tobacco smoke [214]. Research in vitamin C-deficient guinea pigs exposed to tobacco smoke has indicated that vitamin C can protect against protein damage and lipid peroxidation [213,215]. In passive smokers exposed to environmental tobacco smoke, vitamin C supplementation significantly reduced plasma F₂-isoprostane concentrations, a measure of oxidative stress [216]. Tobacco use increases susceptibility to bacterial and viral infections [217,218], in which vitamin C may play a role. For example, in a population-based study the risk of developing obstructive airways disease was significantly higher in those with the lowest plasma vitamin C concentrations (26 µmol/L) compared to never smokers, a risk that decreased with increasing vitamin C concentration [219].

Individuals with diabetes are at greater risk of common infections, including influenza, pneumonia, and foot infections, which are associated with increased morbidity and mortality [220,221]. Several immune-related changes are observed in obesity that contribute towards the development of type 2 diabetes. A major factor is persistent low-grade inflammation of adipose tissue in obese subjects, which plays a role in the progression to insulin resistance and type 2 diabetes, and which is not present in the adipose tissue of lean subjects [222,223]. The adipose tissue is infiltrated by pro-inflammatory macrophages and T-cells, leading to the accumulation of pro-inflammatory cytokines such as interleukins and TNF-α [224,225]. A decrease in plasma vitamin C levels has been observed in studies of type 2 diabetes [18,226], and a major cause of increased need for vitamin C in type 2 diabetes is thought to be the high level of oxidative stress caused by hyperglycemia [10,227,228]. Inverse correlations have been reported between plasma vitamin C concentrations and the risk of diabetes, hemoglobin A1c concentrations (an index of glucose tolerance), fasting and postprandial blood glucose, and oxidative stress [219,229–232]. Meta-analysis of interventional studies has indicated that supplementation with vitamin C can improve glycemic control in type 2 diabetes [233].

Elderly people are particularly susceptible to infections due to immunosenescence and decreased immune cell function [234]. For example, common viral infections such as respiratory illnesses, that are usually self-limiting in healthy young people, can lead to the development of complications such as pneumonia, resulting in increased morbidity and mortality in elderly people. A lower mean vitamin C status has been observed in free-living or institutionalized elderly people, indicated by lowered plasma and leukocyte concentrations [10,235,236], which is of concern because low vitamin C concentrations (<17 µmol/L) in older people (aged 75–82 years) are strongly predictive of all-cause mortality [237]. Acute and chronic diseases that are prevalent in this age group may also play an important part in the reduction of vitamin C reserves [238–240]. Institutionalization in particular is an aggravating factor in this age group, resulting in even lower plasma vitamin C levels than in non-institutionalized elderly people. It is noteworthy that elderly hospitalized patients with acute respiratory infections

have been shown to fare significantly better with vitamin C supplementation than those not receiving the vitamin [241]. Decreased immunological surveillance in individuals older than 60 years also results in greater risk of cancer, and patients with cancer, particularly those undergoing cancer treatments, have compromised immune systems, decreased vitamin C status, and enhanced risk of developing sepsis [242,243]. Hospitalized patients, in general, have lower vitamin C status than the general population [244].

5. Vitamin C and Infection

A major symptom of the vitamin C deficiency disease scurvy is the marked susceptibility to infections, particularly of the respiratory tract, with pneumonia being one of the most frequent complications of scurvy and a major cause of death [7]. Patients with acute respiratory infections, such as pulmonary tuberculosis and pneumonia, have decreased plasma vitamin C concentrations relative to control subjects [245]. Administration of vitamin C to patients with acute respiratory infections returns their plasma vitamin C levels to normal and ameliorates the severity of the respiratory symptoms [246]. Cases of acute lung infections have shown rapid clearance of chest X-rays following administration of intravenous vitamin C [247,248]. This vitamin C-dependent clearance of neutrophils from infected lungs could conceivably be due to enhanced apoptosis and subsequent phagocytosis and clearance of the spent neutrophils by macrophages [73]. Pre-clinical studies of animals with sepsis-induced lung injury have indicated that vitamin C administration can increase alveolar fluid clearance, enhance bronchoalveolar epithelial barrier function, and attenuate sequestration of neutrophils [74], all essential factors for normal lung function.

Meta-analysis has indicated that vitamin C supplementation with doses of 200 mg or more daily is effective in ameliorating the severity and duration of the common cold, and the incidence of the common cold if also exposed to physical stress [249]. Supplementation of individuals who had an inadequate vitamin C status (i.e., $<45 \mu\text{mol/L}$) also decreased the incidence of the common cold [203]. Surprisingly, few studies have assessed vitamin C status during the common cold [250]. Significant decreases in both leukocyte vitamin C levels, and urinary excretion of the vitamin, have been reported to occur during common cold episodes, with levels returning to normal following the infection [251–254]. These changes indicate that vitamin C is utilized during the common cold infection. Administration of gram doses of vitamin C during the common cold episode ameliorated the decline in leukocyte vitamin C levels, suggesting that administration of vitamin C may be beneficial for the recovery process [251].

Beneficial effects of vitamin C on recovery have been noted in pneumonia. In elderly people hospitalized because of pneumonia, who were determined to have very low vitamin C levels, administration of vitamin C reduced the respiratory symptom score in the more severe patients [246]. In other pneumonia patients, low-dose vitamin C (0.25–0.8 g/day) reduced the hospital stay by 19% compared with no vitamin C supplementation, whereas the higher-dose group (0.5–1.6 g/day) reduced the duration by 36% [255]. There was also a positive effect on the normalization of chest X-ray, temperature, and erythrocyte sedimentation rate [255]. Since prophylactic vitamin C administration also appears to decrease the risk of developing more serious respiratory infections, such as pneumonia [256], it is likely that the low vitamin C levels observed during respiratory infections are both a cause and a consequence of the disease.

6. Conclusions

Overall, vitamin C appears to exert a multitude of beneficial effects on cellular functions of both the innate and adaptive immune system. Although vitamin C is a potent antioxidant protecting the body against endogenous and exogenous oxidative challenges, it is likely that its action as a cofactor for numerous biosynthetic and gene regulatory enzymes plays a key role in its immune-modulating effects. Vitamin C stimulates neutrophil migration to the site of infection, enhances phagocytosis and oxidant generation, and microbial killing. At the same time, it protects host tissue from excessive

damage by enhancing neutrophil apoptosis and clearance by macrophages, and decreasing neutrophil necrosis and NETosis. Thus, it is apparent that vitamin C is necessary for the immune system to mount and sustain an adequate response against pathogens, whilst avoiding excessive damage to the host.

Vitamin C appears to be able to both prevent and treat respiratory and systemic infections by enhancing various immune cell functions. Prophylactic prevention of infection requires dietary vitamin C intakes that provide at least adequate, if not saturating plasma levels (i.e., 100–200 mg/day), which optimize cell and tissue levels. In contrast, treatment of established infections requires significantly higher (gram) doses of the vitamin to compensate for the increased metabolic demand.

Epidemiological studies indicate that hypovitaminosis C is still relatively common in Western populations, and vitamin C deficiency is the fourth leading nutrient deficiency in the United States. Reasons include reduced intake combined with limited body stores. Increased needs occur due to pollution and smoking, fighting infections, and diseases with oxidative and inflammatory components, e.g., type 2 diabetes, etc. Ensuring adequate intake of vitamin C through the diet or via supplementation, especially in groups such as the elderly or in individuals exposed to risk factors for vitamin C insufficiency, is required for proper immune function and resistance to infections.

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References

1. Parkin, J.; Cohen, B. An overview of the immune system. *Lancet* **2001**, *357*, 1777–1789. [[CrossRef](#)]
2. Maggini, S.; Wintergerst, E.S.; Beveridge, S.; Hornig, D.H. Selected vitamins and trace elements support immune function by strengthening epithelial barriers and cellular and humoral immune responses. *Br. J. Nutr.* **2007**, *98*, S29–S35. [[CrossRef](#)] [[PubMed](#)]
3. Webb, A.L.; Villamor, E. Update: Effects of antioxidant and non-antioxidant vitamin supplementation on immune function. *Nutr. Rev.* **2007**, *65*, 181. [[CrossRef](#)] [[PubMed](#)]
4. Burns, J.J. Missing step in man, monkey and guinea pig required for the biosynthesis of L-ascorbic acid. *Nature* **1957**, *180*, 553. [[CrossRef](#)] [[PubMed](#)]
5. Nishikimi, M.; Fukuyama, R.; Muroshima, S.; Shimizu, N.; Yagi, K. Cloning and chromosomal mapping of the human nonfunctional gene for L-gulonono-gamma-lactone oxidase, the enzyme for L-ascorbic acid biosynthesis missing in man. *J. Biol. Chem.* **1994**, *269*, 13685–13688. [[PubMed](#)]
6. Sauberlich, H.E. A history of scurvy and vitamin C. In *Vitamin C in Health and Disease*; Packer, L., Fuchs, J., Eds.; Marcel Dekker: New York, NY, USA, 1997; pp. 1–24.
7. Hemila, H. Vitamin C and Infections. *Nutrients* **2017**, *9*, 339. [[CrossRef](#)] [[PubMed](#)]
8. Carr, A.C.; McCall, C. The role of vitamin C in the treatment of pain: New insights. *J. Transl. Med.* **2017**, *15*, 77. [[CrossRef](#)] [[PubMed](#)]
9. Krebs, H.A. The Sheffield Experiment on the vitamin C requirement of human adults. *Proc. Nutr. Soc.* **1953**, *12*, 237–246. [[CrossRef](#)]
10. Institute of Medicine Panel on Dietary Antioxidants and Related Compounds. *Dietary Reference Intakes for Vitamin C, Vitamin E, Selenium, and Carotenoids*; National Academies Press: Washington, DC, USA, 2000.
11. Levine, M.; Dhariwal, K.R.; Welch, R.W.; Wang, Y.; Park, J.B. Determination of optimal vitamin C requirements in humans. *Am. J. Clin. Nutr.* **1995**, *62*, 1347S–1356S. [[PubMed](#)]
12. Carr, A.C.; Frei, B. Toward a new recommended dietary allowance for vitamin C based on antioxidant and health effects in humans. *Am. J. Clin. Nutr.* **1999**, *69*, 1086–1087. [[PubMed](#)]

13. Schleicher, R.L.; Carroll, M.D.; Ford, E.S.; Lacher, D.A. Serum vitamin C and the prevalence of vitamin C deficiency in the United States: 2003–2004 National Health and Nutrition Examination Survey (NHANES). *Am. J. Clin. Nutr.* **2009**, *90*, 1252–1263. [CrossRef] [PubMed]
14. US Centers for Disease Control and Prevention. *Second National Report on Biochemical Indicators of Diet and Nutrition in the US Population 2012*; National Center for Environmental Health: Atlanta, GA, USA, 2012.
15. Maggini, S.; Beveridge, S.; Sorbara, J.; Senatore, G. Feeding the immune system: The role of micronutrients in restoring resistance to infections. *CAB Rev.* **2008**, *3*, 1–21. [CrossRef]
16. Huskisson, E.; Maggini, S.; Ruf, M. The role of vitamins and minerals in energy metabolism and well-being. *J. Int. Med. Res.* **2007**, *35*, 277–289. [CrossRef] [PubMed]
17. Carr, A.; Frei, B. Does vitamin C act as a pro-oxidant under physiological conditions? *FASEB J.* **1999**, *13*, 1007–1024. [PubMed]
18. Mandl, J.; Szarka, A.; Barnhgyi, G. Vitamin C: Update on physiology and pharmacology. *Br. J. Pharmacol.* **2009**, *157*, 1097–1110. [CrossRef] [PubMed]
19. Englard, S.; Seifter, S. The biochemical functions of ascorbic acid. *Annu. Rev. Nutr.* **1986**, *6*, 365–406. [CrossRef] [PubMed]
20. Carr, A.C.; Shaw, G.M.; Fowler, A.A.; Natarajan, R. Ascorbate-dependent vasopressor synthesis: A rationale for vitamin C administration in severe sepsis and septic shock? *Crit. Care* **2015**, *19*, e418. [CrossRef] [PubMed]
21. Kuiper, C.; Vissers, M.C. Ascorbate as a co-factor for Fe- and 2-oxoglutarate dependent dioxygenases: Physiological activity in tumor growth and progression. *Front. Oncol.* **2014**, *4*, 359. [CrossRef] [PubMed]
22. Young, J.I.; Zuchner, S.; Wang, G. Regulation of the epigenome by vitamin C. *Annu. Rev. Nutr.* **2013**, *35*, 545–564. [CrossRef] [PubMed]
23. Pullar, J.M.; Carr, A.C.; Vissers, M.C.M. The roles of vitamin C in skin health. *Nutrients* **2017**, *9*, 866. [CrossRef] [PubMed]
24. Rhie, G.; Shin, M.H.; Seo, J.Y.; Choi, W.W.; Cho, K.H.; Kim, K.H.; Park, K.C.; Eun, H.C.; Chung, J.H. Aging- and photoaging-dependent changes of enzymic and nonenzymic antioxidants in the epidermis and dermis of human skin in vivo. *J. Invest. Dermatol.* **2001**, *117*, 1212–1217. [CrossRef] [PubMed]
25. Shindo, Y.; Witt, E.; Han, D.; Epstein, W.; Packer, L. Enzymic and non-enzymic antioxidants in epidermis and dermis of human skin. *J. Invest. Dermatol.* **1994**, *102*, 122–124. [CrossRef] [PubMed]
26. McArdle, F.; Rhodes, L.E.; Parslew, R.; Jack, C.I.; Friedmann, P.S.; Jackson, M.J. UVR-induced oxidative stress in human skin in vivo: Effects of oral vitamin C supplementation. *Free Radic. Biol. Med.* **2002**, *33*, 1355–1362. [CrossRef]
27. Stelling, H.; Longet, K.; Moodycliffe, A.; Mansourian, R.; Bertschy, E.; Smola, H.; Mauch, C.; Williamson, G. Sodium-dependent vitamin C transporter isoforms in skin: Distribution, kinetics, and effect of UVB-induced oxidative stress. *Free Radic. Biol. Med.* **2007**, *43*, 752–762. [CrossRef] [PubMed]
28. Hodges, R.E.; Baker, E.M.; Hood, J.; Sauberlich, H.E.; March, S.C. Experimental scurvy in man. *Am. J. Clin. Nutr.* **1969**, *22*, 535–548. [PubMed]
29. Hodges, R.E.; Hood, J.; Canham, J.E.; Sauberlich, H.E.; Baker, E.M. Clinical manifestations of ascorbic acid deficiency in man. *Am. J. Clin. Nutr.* **1971**, *24*, 432–443. [PubMed]
30. Kivirikko, K.I.; Myllylä, R.; Pihlajaniemi, T. Protein hydroxylation: Prolyl 4-hydroxylase, an enzyme with four cosubstrates and a multifunctional subunit. *FASEB J.* **1989**, *3*, 1609–1617. [PubMed]
31. Geesin, J.C.; Darr, D.; Kaufman, R.; Murad, S.; Pinnell, S.R. Ascorbic acid specifically increases type I and type III procollagen messenger RNA levels in human skin fibroblast. *J. Invest. Dermatol.* **1988**, *90*, 420–424. [CrossRef] [PubMed]
32. Kishimoto, Y.; Saito, N.; Kurita, K.; Shimokado, K.; Maruyama, N.; Ishigami, A. Ascorbic acid enhances the expression of type I and type 4 collagen and SVCT2 in cultured human skin fibroblasts. *Biochem. Biophys. Res. Commun.* **2013**, *430*, 579–584. [CrossRef] [PubMed]
33. Nausgens, B.V.; Humbert, P.; Rougier, A.; Colige, A.C.; Haftek, M.; Lambert, C.A.; Richard, A.; Creidi, P.; Lapiere, C.M. Topically applied vitamin C enhances the mRNA level of collagens I and III, their processing enzymes and tissue inhibitor of matrix metalloproteinase 1 in the human dermis. *J. Invest. Dermatol.* **2001**, *116*, 853–859. [CrossRef] [PubMed]
34. Tajima, S.; Pinnell, S.R. Ascorbic acid preferentially enhances type I and III collagen gene transcription in human skin fibroblasts. *J. Dermatol. Sci.* **1996**, *11*, 250–253. [CrossRef]

35. Davidson, J.M.; LaValle, P.A.; Zsila, O.; Quaglini, D., Jr.; Giro, M. Ascorbate differentially regulates elastin and collagen biosynthesis in vascular smooth muscle cells and skin fibroblasts by pretranslational mechanisms. *J. Biol. Chem.* **1997**, *272*, 345–352. [\[CrossRef\]](#) [\[PubMed\]](#)
36. Fuchs, J.; Kern, H. Modulation of UV-light-induced skin inflammation by D-alpha-tocopherol and L-ascorbic acid: A clinical study using solar simulated radiation. *Free Radic. Biol. Med.* **1998**, *25*, 1006–1012. [\[CrossRef\]](#)
37. Lauer, A.C.; Groth, N.; Haag, S.F.; Darvin, M.E.; Lademann, J.; Meinke, M.C. Dose-dependent vitamin C uptake and radical scavenging activity in human skin measured with in vivo electron paramagnetic resonance spectroscopy. *Skin Pharmacol. Physiol.* **2013**, *26*, 147–154. [\[CrossRef\]](#) [\[PubMed\]](#)
38. Valacchi, G.; Sticcozzi, C.; Belmonte, G.; Cervellati, F.; Denaude, J.; Chen, N.; Krol, Y.; Oresajo, C. Vitamin C compound mixtures prevent ozone-induced oxidative damage in human keratinocytes as initial assessment of pollution protection. *PLoS ONE* **2015**, *10*, e0131097. [\[CrossRef\]](#) [\[PubMed\]](#)
39. Valacchi, G.; Muresan, X.M.; Sticcozzi, C.; Belmonte, G.; Pecorelli, A.; Cervellati, F.; Denaude, J.; Krol, Y.; Oresajo, C. Ozone-induced damage in 3D-skin model is prevented by topical vitamin C and vitamin E compound mixtures application. *J. Dermatol. Sci.* **2016**, *82*, 209–212. [\[CrossRef\]](#) [\[PubMed\]](#)
40. Lin, J.Y.; Selim, M.A.; Shea, C.R.; Grichnik, J.M.; Omar, M.M.; Monteiro-Riviere, N.A.; Pinnell, S.R. UV photoprotection by combination topical antioxidants vitamin C and vitamin E. *J. Am. Acad. Dermatol.* **2003**, *48*, 866–874. [\[CrossRef\]](#) [\[PubMed\]](#)
41. Pasanen-Seppanen, S.; Suhonen, T.M.; Kirjavainen, M.; Suikko, E.; Urtti, A.; Miettinen, M.; Hytinen, M.; Tammi, M.; Tammi, R. Vitamin C enhances differentiation of a continuous keratinocyte cell line (REK) into epidermis with normal stratum corneum ultrastructure and functional permeability barrier. *Histochem. Cell Biol.* **2001**, *116*, 287–297. [\[CrossRef\]](#) [\[PubMed\]](#)
42. Savini, I.; Catani, M.V.; Rossi, A.; Duranti, G.; Melino, G.; Avigliano, L. Characterization of keratinocyte differentiation induced by ascorbic acid: Protein kinase C involvement and vitamin C homeostasis. *J. Invest. Dermatol.* **2002**, *118*, 372–379. [\[CrossRef\]](#) [\[PubMed\]](#)
43. Poncet, M.; Weerheim, A.; Kempenaar, J.; Mulder, A.; Gooris, G.S.; Bouwstra, J.; Mommaas, A.M. The formation of competent barrier lipids in reconstructed human epidermis requires the presence of vitamin C. *J. Invest. Dermatol.* **1997**, *109*, 348–355. [\[CrossRef\]](#) [\[PubMed\]](#)
44. Uchida, Y.; Behne, M.; Quicc, D.; Elias, P.M.; Holleran, W.M. Vitamin C stimulates sphingolipid production and markers of barrier formation in submerged human keratinocyte cultures. *J. Invest. Dermatol.* **2001**, *117*, 1307–1313. [\[CrossRef\]](#) [\[PubMed\]](#)
45. Kim, K.P.; Shin, K.O.; Park, K.; Yun, H.J.; Mann, S.; Lee, Y.M.; Cho, Y. Vitamin C stimulates epidermal ceramide production by regulating its metabolic enzymes. *Biomol. Ther.* **2015**, *23*, 525–530. [\[CrossRef\]](#) [\[PubMed\]](#)
46. Mohammed, B.M.; Fisher, B.J.; Krskauskas, D.; Ward, S.; Wayne, J.S.; Brophy, D.F.; Fowler, A.A., III; Yager, D.R.; Natarajan, R. Vitamin C promotes wound healing through novel pleiotropic mechanisms. *Int. Wound J.* **2016**, *13*, 572–584. [\[CrossRef\]](#) [\[PubMed\]](#)
47. Duarte, T.L.; Cooke, M.S.; Jones, G.D. Gene expression profiling reveals new protective roles for vitamin C in human skin cells. *Free Radic. Biol. Med.* **2009**, *46*, 78–87. [\[CrossRef\]](#) [\[PubMed\]](#)
48. Desneves, K.J.; Todorovic, B.E.; Cassat, A.; Crowe, T.C. Treatment with supplementary arginine, vitamin C and zinc in patients with pressure ulcers: A randomised controlled trial. *Clin. Nutr.* **2005**, *24*, 979–987. [\[CrossRef\]](#) [\[PubMed\]](#)
49. Taylor, T.V.; Rimmer, S.; Day, B.; Butcher, J.; Dymock, I.W. Ascorbic acid supplementation in the treatment of pressure-sores. *Lancet* **1974**, *2*, 544–546. [\[CrossRef\]](#)
50. Stankova, L.; Gerhardt, N.B.; Nagel, L.; Bigley, R.H. Ascorbate and phagocyte function. *Infect. Immun.* **1975**, *12*, 252–256. [\[PubMed\]](#)
51. Winterbourn, C.C.; Visser, M.C. Changes in ascorbate levels on stimulation of human neutrophils. *Biochim. Biophys. Acta* **1983**, *763*, 175–179. [\[CrossRef\]](#)
52. Parker, A.; Cuddihy, S.L.; Son, T.G.; Visser, M.C.; Winterbourn, C.C. Roles of superoxide and myeloperoxidase in ascorbate oxidation in stimulated neutrophils and H₂O₂-treated HL60 cells. *Free Radic. Biol. Med.* **2011**, *51*, 1399–1405. [\[CrossRef\]](#) [\[PubMed\]](#)
53. Obermayer, H.; Glatthaar, B.; Moser, U.; Schmidt, K.H. Effect of functional stimulation on ascorbate content in phagocytes under physiological and pathological conditions. *Int. Arch. Allergy Appl. Immunol.* **1986**, *81*, 46–50. [\[CrossRef\]](#) [\[PubMed\]](#)

54. Goldschmidt, M.C. Reduced bactericidal activity in neutrophils from scorbutic animals and the effect of ascorbic acid on these target bacteria in vivo and in vitro. *Am. J. Clin. Nutr.* **1991**, *54*, 1214S–1220S. [\[PubMed\]](#)
55. Goldschmidt, M.C.; Masin, W.J.; Brown, L.R.; Wyde, P.R. The effect of ascorbic acid deficiency on leukocyte phagocytosis and killing of *actinomyces viscosus*. *Int. J. Vitam. Nutr. Res.* **1988**, *58*, 326–334. [\[PubMed\]](#)
56. Johnston, C.S.; Huang, S.N. Effect of ascorbic acid nutrition on blood histamine and neutrophil chemotaxis in guinea pigs. *J. Nutr.* **1991**, *121*, 126–130. [\[PubMed\]](#)
57. Rebora, A.; Dallegrì, F.; Patrone, F. Neutrophil dysfunction and repeated infections: Influence of levamisole and ascorbic acid. *Br. J. Dermatol.* **1980**, *102*, 49–56. [\[CrossRef\]](#) [\[PubMed\]](#)
58. Patrone, F.; Dallegrì, F.; Borvini, E.; Minervini, F.; Sacchetti, C. Disorders of neutrophil function in children with recurrent pyogenic infections. *Med. Microbiol. Immunol.* **1982**, *171*, 113–122. [\[CrossRef\]](#) [\[PubMed\]](#)
59. Boura, P.; Tsapas, G.; Papadopoulou, A.; Magoula, I.; Kountouras, G. Monocyte locomotion in anergic chronic brucellosis patients: The in vivo effect of ascorbic acid. *Immunopharmacol. Immunotoxicol.* **1989**, *11*, 119–129. [\[CrossRef\]](#) [\[PubMed\]](#)
60. Anderson, R.; Theron, A. Effects of ascorbate on leucocytes: Part III. In vitro and in vivo stimulation of abnormal neutrophil motility by ascorbate. *S. Afr. Med. J.* **1979**, *56*, 429–433. [\[PubMed\]](#)
61. Johnston, C.S.; Martin, L.J.; Cai, X. Antihistamine effect of supplemental ascorbic acid and neutrophil chemotaxis. *J. Am. Coll. Nutr.* **1992**, *11*, 172–176. [\[PubMed\]](#)
62. Anderson, R.; Oosthuizen, R.; Martitz, R.; Theron, A.; Van Rensburg, A.J. The effects of increasing weekly doses of ascorbate on certain cellular and humoral immune functions in normal volunteers. *Am. J. Clin. Nutr.* **1980**, *33*, 71–76. [\[PubMed\]](#)
63. Anderson, R. Ascorbate-mediated stimulation of neutrophil motility and lymphocyte transformation by inhibition of the peroxidase/H₂O₂/halide system in vitro and in vivo. *Am. J. Clin. Nutr.* **1981**, *34*, 1906–1911. [\[PubMed\]](#)
64. Ganguly, R.; Durieux, M.F.; Waldman, R.H. Macrophage function in vitamin C-deficient guinea pigs. *Am. J. Clin. Nutr.* **1976**, *29*, 762–765. [\[PubMed\]](#)
65. Corberand, J.; Nguyen, F.; Frayse, B.; Enjalbert, L. Malignant external otitis and polymorphonuclear leukocyte migration impairment. Improvement with ascorbic acid. *Arch. Otolaryngol.* **1982**, *108*, 122–124. [\[CrossRef\]](#) [\[PubMed\]](#)
66. Levy, R.; Schläeffer, F. Successful treatment of a patient with recurrent furunculosis by vitamin C: Improvement of clinical course and of impaired neutrophil functions. *Int. J. Dermatol.* **1993**, *32*, 832–834. [\[CrossRef\]](#) [\[PubMed\]](#)
67. Levy, R.; Shriker, O.; Porath, A.; Riesenber, K.; Schläeffer, F. Vitamin C for the treatment of recurrent furunculosis in patients with impaired neutrophil functions. *J. Infect. Dis.* **1996**, *173*, 1502–1505. [\[CrossRef\]](#) [\[PubMed\]](#)
68. Nungesser, W.J.; Ames, A.M. The relationship between ascorbic acid and phagocytic activity. *J. Infect. Dis.* **1948**, *83*, 50–54. [\[CrossRef\]](#) [\[PubMed\]](#)
69. Shilotti, P.G. Phagocytosis and leukocyte enzymes in ascorbic acid deficient guinea pigs. *J. Nutr.* **1977**, *107*, 1513–1516. [\[PubMed\]](#)
70. Shilotti, P.G. Glycolytic, hexose monophosphate shunt and bactericidal activities of leukocytes in ascorbic acid deficient guinea pigs. *J. Nutr.* **1977**, *107*, 1507–1512. [\[PubMed\]](#)
71. Sharma, P.; Raghavan, S.A.; Saini, R.; Dikshit, M. Ascorbate-mediated enhancement of reactive oxygen species generation from polymorphonuclear leukocytes: Modulatory effect of nitric oxide. *J. Leukoc. Biol.* **2004**, *75*, 1070–1078. [\[CrossRef\]](#) [\[PubMed\]](#)
72. Rebora, A.; Crovato, F.; Dallegrì, F.; Patrone, F. Repeated staphylococcal pyoderma in two siblings with defective neutrophil bacterial killing. *Dermatologica* **1980**, *160*, 106–112. [\[CrossRef\]](#) [\[PubMed\]](#)
73. Vissers, M.C.; Wilkie, R.P. Ascorbate deficiency results in impaired neutrophil apoptosis and clearance and is associated with up-regulation of hypoxia-inducible factor 1 α . *J. Leukoc. Biol.* **2007**, *81*, 1236–1244. [\[CrossRef\]](#) [\[PubMed\]](#)
74. Fisher, B.J.; Kraskauskas, D.; Martin, E.J.; Farkas, D.; Wegelin, J.A.; Brophy, D.; Ward, K.R.; Voelkel, N.F.; Fowler, A.A., III; Natarajan, R. Mechanisms of attenuation of abdominal sepsis induced acute lung injury by ascorbic acid. *Am. J. Physiol. Lung Cell. Mol. Physiol.* **2012**, *303*, L20–L32. [\[CrossRef\]](#) [\[PubMed\]](#)

75. Mohammed, B.M.; Fisher, B.J.; Kraskauskas, D.; Farkas, D.; Brophy, D.F.; Fowler, A.A.; Natarajan, R. Vitamin C: A novel regulator of neutrophil extracellular trap formation. *Nutrients* **2013**, *5*, 3131–3151. [\[CrossRef\]](#) [\[PubMed\]](#)
76. Huijskens, M.J.; Walczak, M.; Koller, N.; Briede, J.J.; Senden-Gijbers, B.L.; Schrijderberg, M.C.; Bos, G.M.; Germeeraad, W.T. Technical advance: Ascorbic acid induces development of double-positive T cells from human hematopoietic stem cells in the absence of stromal cells. *J. Leukoc. Biol.* **2014**, *96*, 1165–1175. [\[CrossRef\]](#) [\[PubMed\]](#)
77. Molina, N.; Morandi, A.C.; Bolin, A.F.; Otton, R. Comparative effect of fucoxanthin and vitamin C on oxidative and functional parameters of human lymphocytes. *Int. Immunopharmacol.* **2014**, *22*, 41–50. [\[CrossRef\]](#) [\[PubMed\]](#)
78. Tanaka, M.; Muto, N.; Gohda, E.; Yamamoto, I. Enhancement by ascorbic acid 2-glucoside or repeated additions of ascorbate of mitogen-induced IgM and IgG productions by human peripheral blood lymphocytes. *Jpn. J. Pharmacol.* **1994**, *66*, 451–456. [\[CrossRef\]](#) [\[PubMed\]](#)
79. Manning, J.; Mitchell, B.; Appadurai, D.A.; Shukya, A.; Pierce, L.J.; Wang, H.; Nganga, V.; Swanson, P.C.; May, J.M.; Tantin, D.; et al. Vitamin C promotes maturation of T-cells. *Antioxid. Redox Signal.* **2013**, *19*, 2054–2067. [\[CrossRef\]](#) [\[PubMed\]](#)
80. Kennes, B.; Dumont, I.; Brohee, D.; Hubert, C.; Neve, P. Effect of vitamin C supplements on cell-mediated immunity in old people. *Gerontology* **1983**, *29*, 305–310. [\[CrossRef\]](#) [\[PubMed\]](#)
81. Anderson, R.; Hay, L.; van Wyk, H.; Oosthuizen, R.; Theron, A. The effect of ascorbate on cellular humoral immunity in asthmatic children. *S. Afr. Med. J.* **1980**, *58*, 974–977. [\[PubMed\]](#)
82. Fraser, R.C.; Pavlovic, S.; Kurahara, C.G.; Murata, A.; Peterson, N.S.; Taylor, K.B.; Feigen, G.A. The effect of variations in vitamin C intake on the cellular immune response of guinea pigs. *Am. J. Clin. Nutr.* **1980**, *33*, 839–847. [\[PubMed\]](#)
83. Feigen, G.A.; Smith, B.H.; Dix, C.E.; Flynn, C.J.; Peterson, N.S.; Rosenberg, L.T.; Pavlovic, S.; Leibovitz, B. Enhancement of antibody production and protection against systemic anaphylaxis by large doses of vitamin C. *Res. Commun. Chem. Pathol. Pharmacol.* **1982**, *38*, 313–333. [\[CrossRef\]](#)
84. Prinz, W.; Bloch, J.; Gilich, G.; Mitchell, G. A systematic study of the effect of vitamin C supplementation on the humoral immune response in ascorbate-dependent mammals. I. The antibody response to sheep red blood cells (a T-dependent antigen) in guinea pigs. *Int. J. Vitam. Nutr. Res.* **1980**, *50*, 294–300. [\[PubMed\]](#)
85. Prinz, W.; Bortz, R.; Bregin, B.; Hersch, M. The effect of ascorbic acid supplementation on some parameters of the human immunological defence system. *Int. J. Vitam. Nutr. Res.* **1977**, *47*, 248–257. [\[PubMed\]](#)
86. Chen, Y.; Luo, G.; Yuan, J.; Wang, Y.; Yang, X.; Wang, X.; Li, G.; Liu, Z.; Zhong, N. Vitamin C mitigates oxidative stress and tumor necrosis factor- α in severe community-acquired pneumonia and LPS-induced macrophages. *Mediators Inflamm.* **2014**, *2014*, 426740. [\[CrossRef\]](#) [\[PubMed\]](#)
87. Jeng, K.C.; Yang, C.S.; Sui, W.Y.; Tsai, Y.S.; Liao, W.J.; Kuo, J.S. Supplementation with vitamins C and E enhances cytokine production by peripheral blood mononuclear cells in healthy adults. *Am. J. Clin. Nutr.* **1996**, *64*, 960–965. [\[PubMed\]](#)
88. Kim, Y.; Kim, H.; Bae, S.; Choi, J.; Lim, S.Y.; Lee, N.; Korg, J.M.; Hwang, Y.I.; Kang, J.S.; Lee, W.J. Vitamin C is an essential factor on the anti-viral immune responses through the production of interferon- α /b at the initial stage of influenza A virus (H₃N₂) infection. *Immune Netw.* **2013**, *13*, 70–74. [\[CrossRef\]](#) [\[PubMed\]](#)
89. Gao, Y.L.; Lu, B.; Zhai, J.H.; Liu, Y.C.; Qi, H.X.; Yao, Y.; Chai, Y.F.; Shou, S.T. The parenteral vitamin C improves sepsis and sepsis-induced multiple organ dysfunction syndrome via preventing cellular immunosuppression. *Mediat. Inflamm.* **2017**, *2017*, 4024672. [\[CrossRef\]](#) [\[PubMed\]](#)
90. Portugal, C.C.; Socolato, R.; Canedo, T.; Silva, C.M.; Martins, T.; Correias, V.S.; Loiola, E.C.; Gess, B.; Rohrer, D.; Santiago, A.R.; et al. Caveolin-1-mediated internalization of the vitamin C transporter SVCT2 in microglia triggers an inflammatory phenotype. *Sci. Signal.* **2017**, *10*. [\[CrossRef\]](#) [\[PubMed\]](#)
91. Dahl, H.; Degre, M. The effect of ascorbic acid on production of human interferon and the antiviral activity in vitro. *Acta Pathol. Microbiol. Scand. B* **1976**, *84b*, 280–284. [\[CrossRef\]](#) [\[PubMed\]](#)
92. Karpinska, T.; Kawecky, Z.; Kandefer-Szerszen, M. The influence of ultraviolet irradiation, L-ascorbic acid and calcium chloride on the induction of interferon in human embryo fibroblasts. *Arch. Immunol. Ther. Exp.* **1982**, *30*, 33–37.
93. Siegel, B.V. Enhancement of interferon production by poly(rI)-poly(rC) in mouse cell cultures by ascorbic acid. *Nature* **1975**, *254*, 531–532. [\[CrossRef\]](#) [\[PubMed\]](#)

94. Canali, R.; Natarrelli, L.; Leoni, G.; Azzini, E.; Comitato, R.; Sancak, O.; Barella, L.; Virgili, F. Vitamin C supplementation modulates gene expression in peripheral blood mononuclear cells specifically upon an inflammatory stimulus: A pilot study in healthy subjects. *Genes Nutr.* **2014**, *9*, 390. [\[CrossRef\]](#) [\[PubMed\]](#)
95. Dawson, W.; West, G.B. The influence of ascorbic acid on histamine metabolism in guinea-pigs. *Br. J. Pharmacol. Chemother.* **1965**, *24*, 725–734. [\[CrossRef\]](#) [\[PubMed\]](#)
96. Nandi, B.K.; Subramanian, N.; Majumder, A.K.; Chatterjee, I.B. Effect of ascorbic acid on detoxification of histamine under stress conditions. *Biochem. Pharmacol.* **1974**, *23*, 643–647. [\[CrossRef\]](#)
97. Subramanian, N.; Nandi, B.K.; Majumder, A.K.; Chatterjee, I.B. Role of L-ascorbic acid on detoxification of histamine. *Biochem. Pharmacol.* **1973**, *22*, 1671–1673. [\[CrossRef\]](#)
98. Chatterjee, I.B.; Gupta, S.D.; Majumder, A.K.; Nandi, B.K.; Subramanian, N. Effect of ascorbic acid on histamine metabolism in scorbutic guinea-pigs. *J. Physiol.* **1975**, *251*, 271–279. [\[CrossRef\]](#) [\[PubMed\]](#)
99. Clemetson, C.A. Histamine and ascorbic acid in human blood. *J. Nutr.* **1980**, *110*, 662–668. [\[PubMed\]](#)
100. Johnston, C.S.; Solomon, R.E.; Corte, C. Vitamin C depletion is associated with alterations in blood histamine and plasma free carnitine in adults. *J. Am. Coll. Nutr.* **1996**, *15*, 586–591. [\[CrossRef\]](#) [\[PubMed\]](#)
101. Hagel, A.F.; Layritz, C.M.; Hagel, W.H.; Hagel, H.J.; Hagel, E.; Dauth, W.; Kressel, J.; Regnet, T.; Rosenberg, A.; Neurath, M.F.; et al. Intravenous infusion of ascorbic acid decreases serum histamine concentrations in patients with allergic and non-allergic diseases. *Naunyn-Schmiedeberg's Arch. Pharmacol.* **2013**, *386*, 789–793. [\[CrossRef\]](#) [\[PubMed\]](#)
102. Bruno, R.S.; Leonard, S.W.; Atkinson, J.; Montine, T.J.; Ramakrishnan, R.; Bray, T.M.; Traber, M.G. Faster plasma vitamin E disappearance in smokers is normalized by vitamin C supplementation. *Free Radic. Biol. Med.* **2006**, *40*, 689–697. [\[CrossRef\]](#) [\[PubMed\]](#)
103. Parsons, K.K.; Maeda, N.; Yamauchi, M.; Barnes, A.J.; Koller, B.H. Ascorbic acid-independent synthesis of collagen in mice. *Am. J. Physiol. Endocrinol. Metab.* **2006**, *290*, E1131–E1139. [\[CrossRef\]](#) [\[PubMed\]](#)
104. Ross, R.; Benditt, E.P. Wound healing and collagen formation. II. Fine structure in experimental scurvy. *J. Cell Biol.* **1962**, *12*, 533–551. [\[CrossRef\]](#) [\[PubMed\]](#)
105. Fukushima, R.; Yamazaki, E. Vitamin C requirement in surgical patients. *Curr. Opin. Clin. Nutr. Metab. Care* **2010**, *13*, 669–676. [\[CrossRef\]](#) [\[PubMed\]](#)
106. Blass, S.C.; Goost, H.; Tolba, R.H.; Stoffel-Wagner, B.; Kabir, K.; Burger, C.; Stehle, P.; Ellinger, S. Time to wound closure in trauma patients with disorders in wound healing is shortened by supplements containing antioxidant micronutrients and glutamine: A PRCT. *Clin. Nutr.* **2012**, *31*, 469–475. [\[CrossRef\]](#) [\[PubMed\]](#)
107. Cereda, E.; Gini, A.; Pedrolli, C.; Vanotti, A. Disease-specific, versus standard, nutritional support for the treatment of pressure ulcers in institutionalized older adults: A randomized controlled trial. *J. Am. Geriatr. Soc.* **2009**, *57*, 1395–1402. [\[CrossRef\]](#) [\[PubMed\]](#)
108. Martin, P.; Leibovich, S.J. Inflammatory cells during wound repair: The good, the bad and the ugly. *Trends Cell Biol.* **2005**, *15*, 599–607. [\[CrossRef\]](#) [\[PubMed\]](#)
109. Wilgus, T.A.; Roy, S.; McDaniel, J.C. Neutrophils and Wound Repair: Positive Actions and Negative Reactions. *Adv. Wound Care* **2013**, *2*, 379–388. [\[CrossRef\]](#) [\[PubMed\]](#)
110. Wong, S.L.; Demers, M.; Martinod, K.; Gallant, M.; Wang, Y.; Goldfine, A.B.; Kahn, C.R.; Wagner, D.D. Diabetes primes neutrophils to undergo NETosis, which impairs wound healing. *Nat. Med.* **2015**, *21*, 815–819. [\[CrossRef\]](#) [\[PubMed\]](#)
111. Washko, P.; Rotrosen, D.; Levine, M. Ascorbic acid transport and accumulation in human neutrophils. *J. Biol. Chem.* **1989**, *264*, 18996–19002. [\[PubMed\]](#)
112. Bergsten, P.; Amitai, G.; Kehr, J.; Dhariwal, K.R.; Klein, H.G.; Levine, M. Millimolar concentrations of ascorbic acid in purified human mononuclear leukocytes. Depletion and reaccumulation. *J. Biol. Chem.* **1990**, *265*, 2584–2587. [\[PubMed\]](#)
113. Evans, R.M.; Currie, L.; Campbell, A. The distribution of ascorbic acid between various cellular components of blood, in normal individuals, and its relation to the plasma concentration. *Br. J. Nutr.* **1982**, *47*, 473–482. [\[CrossRef\]](#) [\[PubMed\]](#)
114. Levine, M.; Conry-Cantilena, C.; Wang, Y.; Welch, R.W.; Washko, P.W.; Dhariwal, K.R.; Park, J.B.; Lazarev, A.; Graumlich, J.F.; King, J.; et al. Vitamin C pharmacokinetics in healthy volunteers: Evidence for a recommended dietary allowance. *Proc. Natl. Acad. Sci. USA* **1996**, *93*, 3704–3709. [\[CrossRef\]](#) [\[PubMed\]](#)
115. Levine, M.; Wang, Y.; Padayatty, S.J.; Morrow, J. A new recommended dietary allowance of vitamin C for healthy young women. *Proc. Natl. Acad. Sci. USA* **2001**, *98*, 9842–9846. [\[CrossRef\]](#) [\[PubMed\]](#)

116. Carr, A.C.; Bozonet, S.M.; Pullar, J.M.; Simcock, J.W.; Visser, M.C. Human skeletal muscle ascorbate is highly responsive to changes in vitamin C intake and plasma concentrations. *Am. J. Clin. Nutr.* **2013**, *97*, 800–807. [\[CrossRef\]](#) [\[PubMed\]](#)
117. Visser, M.C.; Bozonet, S.M.; Pearson, J.F.; Braithwaite, L.J. Dietary ascorbate intake affects steady state tissue concentrations in vitamin C-deficient mice: Tissue deficiency after suboptimal intake and superior bioavailability from a food source (kiwifruit). *Am. J. Clin. Nutr.* **2011**, *93*, 292–301. [\[CrossRef\]](#) [\[PubMed\]](#)
118. Corpe, C.P.; Lee, J.H.; Kwon, O.; Eck, P.; Narayanan, J.; Kirk, K.L.; Levine, M. 6-Bromo-6-deoxy-L-ascorbic acid: An ascorbate analog specific for Na⁺-dependent vitamin C transporter but not glucose transporter pathways. *J. Biol. Chem.* **2005**, *280*, 5211–5220. [\[CrossRef\]](#) [\[PubMed\]](#)
119. Washko, P.W.; Wang, Y.; Levine, M. Ascorbic acid recycling in human neutrophils. *J. Biol. Chem.* **1993**, *268*, 15531–15535. [\[PubMed\]](#)
120. Buettner, G.R. The pecking order of free radicals and antioxidants: Lipid peroxidation, alpha-tocopherol, and ascorbate. *Arch. Biochem. Biophys.* **1993**, *300*, 535–543. [\[CrossRef\]](#) [\[PubMed\]](#)
121. Sen, C.K.; Packer, L. Antioxidant and redox regulation of gene transcription. *FASEB J.* **1996**, *10*, 709–720. [\[PubMed\]](#)
122. Li, N.; Karin, M. Is NF-kappaB the sensor of oxidative stress? *FASEB J.* **1999**, *13*, 1137–1143. [\[PubMed\]](#)
123. Macdonald, J.; Gailey, H.E.; Webster, N.R. Oxidative stress and gene expression in sepsis. *Br. J. Anaesth.* **2003**, *90*, 221–232. [\[CrossRef\]](#) [\[PubMed\]](#)
124. Tan, P.H.; Sagoo, P.; Chan, C.; Yates, J.B.; Campbell, J.; Beutelspacher, S.C.; Foxwell, B.M.; Lombardi, G.; George, A.J. Inhibition of NF-kappa B and oxidative pathways in human dendritic cells by antioxidative vitamins generates regulatory T cells. *J. Immunol.* **2008**, *174*, 7633–7644. [\[CrossRef\]](#) [\[PubMed\]](#)
125. Winterbourn, C.C.; Hampton, M.B. Thiol chemistry and specificity in redox signaling. *Free Radic. Biol. Med.* **2008**, *45*, 549–561. [\[CrossRef\]](#) [\[PubMed\]](#)
126. Griffiths, H.R.; Willets, R.S.; Grant, M.M.; Mistry, N.; Lunec, J.; Bevan, R.J. In vivo vitamin C supplementation increases phosphoinositol transfer protein expression in peripheral blood mononuclear cells from healthy individuals. *Br. J. Nutr.* **2009**, *101*, 1432–1439. [\[CrossRef\]](#) [\[PubMed\]](#)
127. Grant, M.M.; Mistry, N.; Lunec, J.; Griffiths, H.R. Dose-dependent modulation of the T cell proteome by ascorbic acid. *Br. J. Nutr.* **2007**, *97*, 19–26. [\[CrossRef\]](#) [\[PubMed\]](#)
128. Lammernann, T. In the eye of the neutrophil swarm-navigation signals that bring neutrophils together in inflamed and infected tissues. *J. Leukoc. Biol.* **2016**, *100*, 55–63. [\[CrossRef\]](#) [\[PubMed\]](#)
129. Demaret, J.; Venet, F.; Friggeri, A.; Cazalis, M.A.; Plassais, J.; Jallades, L.; Malcus, C.; Poitevin-Later, F.; Textoris, J.; Lepape, A.; et al. Marked alterations of neutrophil functions during sepsis-induced immunosuppression. *J. Leukoc. Biol.* **2015**, *98*, 1081–1090. [\[CrossRef\]](#) [\[PubMed\]](#)
130. Arraes, S.M.; Freitas, M.S.; da Silva, S.V.; de Paula Neto, H.A.; Alves-Filho, J.C.; Auxiliadora Martins, M.; Basile-Filho, A.; Tavares-Murta, B.M.; Barja-Fidalgo, C.; Cunha, F.Q. Impaired neutrophil chemotaxis in sepsis associates with GRK expression and inhibition of actin assembly and tyrosine phosphorylation. *Bloof* **2006**, *108*, 2906–2913. [\[CrossRef\]](#) [\[PubMed\]](#)
131. Chishti, A.D.; Shenton, B.K.; Kirby, J.A.; Baudouin, S.V. Neutrophil chemotaxis and receptor expression in clinical septic shock. *Intensive Care Med.* **2004**, *30*, 605–611. [\[CrossRef\]](#) [\[PubMed\]](#)
132. Tavares-Murta, B.M.; Zapanoli, M.; Ferreira, R.B.; Silva-Vergara, M.L.; Oliveira, C.H.; Murta, E.F.; Ferreira, S.H.; Cunha, F.Q. Failure of neutrophil chemotactic function in septic patients. *Crit. Care Med.* **2002**, *30*, 1056–1061. [\[CrossRef\]](#) [\[PubMed\]](#)
133. Hotchkiss, R.S.; Monneret, G.; Payen, D. Sepsis-induced immunosuppression: From cellular dysfunctions to immunotherapy. *Nat. Rev. Immunol.* **2013**, *13*, 862–874. [\[CrossRef\]](#) [\[PubMed\]](#)
134. Vohra, K.; Khan, A.J.; Telang, V.; Rosenfeld, W.; Evans, H.E. Improvement of neutrophil migration by systemic vitamin C in neonates. *J. Perinatol.* **1990**, *10*, 134–136. [\[PubMed\]](#)
135. Roos, D. Chronic granulomatous disease. *Br. Med. Bull.* **2016**, *118*, 50–63. [\[CrossRef\]](#) [\[PubMed\]](#)
136. Introne, W.; Boissy, R.E.; Gahl, W.A. Clinical, molecular, and cell biological aspects of Chediak-Higashi syndrome. *Mol. Genet. Metab.* **1999**, *68*, 283–303. [\[CrossRef\]](#) [\[PubMed\]](#)
137. Anderson, R.; Dittich, O.C. Effects of ascorbate on leucocytes: Part IV. Increased neutrophil function and clinical improvement after oral ascorbate in 2 patients with chronic granulomatous disease. *S. Afr. Med. J.* **1979**, *56*, 476–480. [\[PubMed\]](#)

138. Anderson, R. Assessment of oral ascorbate in three children with chronic granulomatous disease and defective neutrophil motility over a 2-year period. *Clin. Exp. Immunol.* **1981**, *43*, 180–188. [\[PubMed\]](#)
139. Anderson, R. Effects of ascorbate on normal and abnormal leucocyte functions. *Int. J. Vitam. Nutr. Res. Suppl.* **1982**, *23*, 23–34. [\[PubMed\]](#)
140. Gallin, J.L.; Elin, R.J.; Hubert, R.T.; Fauci, A.S.; Kaliner, M.A.; Wolff, S.M. Efficacy of ascorbic acid in Chediak-Higashi syndrome (CHS): Studies in humans and mice. *Blood* **1979**, *53*, 226–234. [\[PubMed\]](#)
141. Boxer, L.A.; Watanabe, A.M.; Rister, M.; Besch, H.R., Jr.; Allen, J.; Baehner, R.L. Correction of leukocyte function in Chediak-Higashi syndrome by ascorbate. *N. Engl. J. Med.* **1976**, *295*, 1041–1045. [\[CrossRef\]](#) [\[PubMed\]](#)
142. Yegin, O.; Sanal, O.; Yeralan, O.; Gurgey, A.; Berkel, A.I. Defective lymphocyte locomotion in Chediak-Higashi syndrome. *Am. J. Dis. Child.* **1983**, *137*, 771–773. [\[PubMed\]](#)
143. Weening, R.S.; Schoonl, E.P.; Roos, D.; van Schaik, M.L.; Voetman, A.A.; Bot, A.A.; Batenburg-Plenter, A.M.; Willems, C.; Zeijlenaker, W.P.; Astaldi, A. Effect of ascorbate on abnormal neutrophil, platelet and lymphocytic function in a patient with the Chediak-Higashi syndrome. *Blood* **1981**, *57*, 856–865. [\[PubMed\]](#)
144. Boxer, L.A.; Vanderbilt, B.; Bonsib, S.; Jersild, R.; Yang, H.H.; Baehner, R.L. Enhancement of chemotactic response and microtubule assembly in human leukocytes by ascorbic acid. *J. Cell. Physiol.* **1979**, *100*, 119–126. [\[CrossRef\]](#) [\[PubMed\]](#)
145. Boxer, L.A.; Albertini, D.F.; Baehner, R.L.; Oliver, J.M. Impaired microtubule assembly and polymorphonuclear leukocyte function in the Chediak-Higashi syndrome correctable by ascorbic acid. *Br. J. Haematol.* **1979**, *43*, 207–213. [\[CrossRef\]](#) [\[PubMed\]](#)
146. Parker, W.H.; Rhee, E.M.; Qu, Z.C.; Hecker, M.R.; May, J.M. Intracellular ascorbate tightens the endothelial permeability barrier through Epc1 and the tubulin cytoskeleton. *Am. J. Physiol. Cell Physiol.* **2016**, *311*, C652–C662. [\[CrossRef\]](#) [\[PubMed\]](#)
147. Bozonet, S.M.; Carr, A.C.; Pullar, J.M.; Visser, M.C.M. Enhanced human neutrophil vitamin C status, chemotaxis and oxidant generation following dietary supplementation with vitamin C-rich SunGold kiwifruit. *Nutrients* **2015**, *7*, 2574–2588. [\[CrossRef\]](#) [\[PubMed\]](#)
148. De la Fuente, M.; Ferrandez, M.D.; Burgos, M.S.; Soler, A.; Prieto, A.; Miquel, J. Immune function in aged women is improved by ingestion of vitamins C and E. *Can. J. Physiol. Pharmacol.* **1998**, *76*, 373–380. [\[CrossRef\]](#) [\[PubMed\]](#)
149. Winterbourn, C.C.; Kettle, A.J.; Hampton, M.B. Reactive oxygen species and neutrophil function. *Annu. Rev. Biochem.* **2016**, *85*, 765–792. [\[CrossRef\]](#) [\[PubMed\]](#)
150. Patrone, F.; Dallogri, F.; Bonvini, E.; Minervini, F.; Sacchetti, C. Effects of ascorbic acid on neutrophil function. Studies on normal and chronic granulomatous disease neutrophils. *Acta Vitaminol. Enzymol.* **1982**, *4*, 163–168. [\[PubMed\]](#)
151. Forszwar, N.; Lucas, C.F.; Joss, D.V.; Hugh-Jones, K.; Hobbs, J.R. Ascorbate (1 g/day) does not help the phagocyte killing defect of X-linked chronic granulomatous disease. *Clin. Exp. Immunol.* **1983**, *51*, 99–102. [\[PubMed\]](#)
152. Hampton, M.B.; Kettle, A.J.; Winterbourn, C.C. Inside the neutrophil phagosome: Oxidants, myeloperoxidase, and bacterial killing. *Blood* **1998**, *92*, 3007–3017. [\[PubMed\]](#)
153. Wernisch, C.; Graninger, W. Are soluble factors relevant for polymorphonuclear leukocyte dysregulation in septicemia? *Clin. Diagn. Lab. Immunol.* **1995**, *2*, 241–245. [\[PubMed\]](#)
154. Danikas, D.D.; Karakantza, M.; Theodorou, G.L.; Sakellariopoulos, G.C.; Gogos, C.A. Prognostic value of phagocytic activity of neutrophils and monocytes in sepsis. Correlation to CD64 and CD14 antigen expression. *Clin. Exp. Immunol.* **2008**, *154*, 87–97. [\[CrossRef\]](#) [\[PubMed\]](#)
155. Stephan, F.; Yang, K.; Tankovic, J.; Soussy, C.J.; Dhonneur, G.; Duvaldestin, P.; Brochard, L.; Brun-Buisson, C.; Harf, A.; Delclaux, C. Impairment of polymorphonuclear neutrophil functions precedes nosocomial infections in critically ill patients. *Crit. Care Med.* **2002**, *30*, 315–322. [\[CrossRef\]](#) [\[PubMed\]](#)
156. McGovern, N.N.; Cowburn, A.S.; Porter, L.; Walmsley, S.R.; Summers, C.; Thompson, A.A.; Anwar, S.; Wilcocks, L.C.; Whyte, M.K.; Condliffe, A.M.; et al. Hypoxia selectively inhibits respiratory burst activity and killing of *Staphylococcus aureus* in human neutrophils. *J. Immunol.* **2011**, *186*, 453–463. [\[CrossRef\]](#) [\[PubMed\]](#)

157. Drife, G.; Dunn-Siegrist, I.; Tissieres, P.; Pugin, J. Innate immune functions of immature neutrophils in patients with sepsis and severe systemic inflammatory response syndrome. *Crit. Care Med.* **2013**, *41*, 820–832. [\[CrossRef\]](#) [\[PubMed\]](#)
158. Bass, D.A.; Olbrantz, P.; Szejda, P.; Seeds, M.C.; McCall, C.E. Subpopulations of neutrophils with increased oxidative product formation in blood of patients with infection. *J. Immunol.* **1986**, *136*, 860–866. [\[PubMed\]](#)
159. Pillay, J.; Ramakers, B.P.; Kamp, V.M.; Loi, A.L.; Lam, S.W.; Hielbrink, F.; Leenen, L.P.; Tool, A.T.; Pickkers, P.; Koederman, L. Functional heterogeneity and differential priming of circulating neutrophils in human experimental endotoxemia. *J. Leukoc. Biol.* **2010**, *88*, 211–220. [\[CrossRef\]](#) [\[PubMed\]](#)
160. Weniach, C.; Fladerer, P.; Patruta, S.; Krause, R.; Hori, W. Assessment of neutrophil function in patients with septic shock: Comparison of methods. *Clin. Diagn. Lab. Immunol.* **2001**, *8*, 178–180. [\[CrossRef\]](#) [\[PubMed\]](#)
161. Fox, S.; Leitch, A.E.; Duffin, R.; Haslett, C.; Rossi, A.G. Neutrophil apoptosis: Relevance to the innate immune response and inflammatory disease. *J. Innate Immun.* **2010**, *2*, 216–227. [\[CrossRef\]](#) [\[PubMed\]](#)
162. Hampton, M.B.; Fadeel, B.; Orrenius, S. Redox regulation of the caspases during apoptosis. *Ann. N. Y. Acad. Sci.* **1998**, *854*, 328–335. [\[CrossRef\]](#) [\[PubMed\]](#)
163. Fadeel, B.; Ahlin, A.; Henter, J.I.; Orrenius, S.; Hampton, M.B. Involvement of caspases in neutrophil apoptosis: Regulation by reactive oxygen species. *Blood* **1998**, *92*, 4808–4818. [\[PubMed\]](#)
164. Wilkie, R.P.; Visers, M.C.; Draganow, M.; Hampton, M.B. A functional NADPH oxidase prevents caspase involvement in the clearance of phagocytic neutrophils. *Infect. Immun.* **2007**, *75*, 3256–3263. [\[CrossRef\]](#) [\[PubMed\]](#)
165. Keel, M.; Ungethum, U.; Steckholzer, U.; Niederer, E.; Hartung, T.; Trentz, O.; Ertel, W. Interleukin-10 counterregulates proinflammatory cytokine-induced inhibition of neutrophil apoptosis during severe sepsis. *Blood* **1997**, *90*, 3356–3363. [\[PubMed\]](#)
166. Jimenez, M.F.; Watson, R.W.; Parodo, J.; Evans, D.; Foster, D.; Steinberg, M.; Rotstein, O.D.; Marshall, J.C. Dysregulated expression of neutrophil apoptosis in the systemic inflammatory response syndrome. *Arch. Surg.* **1997**, *132*, 1263–1269. [\[CrossRef\]](#) [\[PubMed\]](#)
167. Harter, L.; Mica, L.; Stocker, R.; Trentz, O.; Keel, M. Mcl-1 correlates with reduced apoptosis in neutrophils from patients with sepsis. *J. Am. Coll. Surg.* **2003**, *197*, 964–973. [\[CrossRef\]](#) [\[PubMed\]](#)
168. Taneja, R.; Parodo, J.; Jia, S.H.; Kapus, A.; Rotstein, O.D.; Marshall, J.C. Delayed neutrophil apoptosis in sepsis is associated with maintenance of mitochondrial transmembrane potential and reduced caspase-9 activity. *Crit. Care Med.* **2004**, *32*, 1460–1469. [\[CrossRef\]](#) [\[PubMed\]](#)
169. Fotouhi-Ardakani, N.; Kebir, D.E.; Pierre-Charles, N.; Wang, L.; Ahern, S.P.; Filep, J.G.; Milot, E. Role for myeloid nuclear differentiation antigen in the regulation of neutrophil apoptosis during sepsis. *Am. J. Respir. Crit. Care Med.* **2010**, *182*, 341–350. [\[CrossRef\]](#) [\[PubMed\]](#)
170. Paunel-Gorgulu, A.; Flohe, S.; Scholz, M.; Windolf, J.; Logters, T. Increased serum soluble Fas after major trauma is associated with delayed neutrophil apoptosis and development of sepsis. *Crit. Care* **2011**, *15*, R20. [\[CrossRef\]](#) [\[PubMed\]](#)
171. Paunel-Gorgulu, A.; Kirichevska, T.; Logters, T.; Windolf, J.; Flohe, S. Molecular mechanisms underlying delayed apoptosis in neutrophils from multiple trauma patients with and without sepsis. *Mol. Med.* **2012**, *18*, 325–335. [\[CrossRef\]](#) [\[PubMed\]](#)
172. Tamayo, E.; Gomez, E.; Bustamante, J.; Gomez-Herreras, J.I.; Fonteriz, R.; Bobillo, F.; Bermejo-Martin, J.F.; Castrodeza, J.; Heredia, M.; Fierro, I.; et al. Evolution of neutrophil apoptosis in septic shock survivors and nonsurvivors. *J. Crit. Care* **2012**, *27*, 415. [\[CrossRef\]](#) [\[PubMed\]](#)
173. Falkow, L.; Fochesatto Filho, L.; Bozzetti, M.C.; Milani, A.R.; Rodrigues Filho, E.M.; Ladniuk, R.M.; Pierozan, P.; de Moura, R.M.; Prolla, J.C.; Vachon, E.; et al. Neutrophil apoptosis: A marker of disease severity in sepsis and sepsis-induced acute respiratory distress syndrome. *Crit. Care* **2006**, *10*, R155. [\[CrossRef\]](#) [\[PubMed\]](#)
174. Ertel, W.; Keel, M.; Infanger, M.; Ungethum, U.; Steckholzer, U.; Trentz, O. Circulating mediators in serum of injured patients with septic complications inhibit neutrophil apoptosis through up-regulation of protein-tyrosine phosphorylation. *J. Trauma* **1998**, *44*, 767–775. [\[CrossRef\]](#) [\[PubMed\]](#)
175. Parlato, M.; Souza-Fonseca-Guimaraes, F.; Philippart, F.; Misset, B.; Adib-Conquy, M.; Cavaillon, J.M. CD24-triggered caspase-dependent apoptosis via mitochondrial membrane depolarization and reactive oxygen species production of human neutrophils is impaired in sepsis. *J. Immunol.* **2014**, *192*, 2449–2459. [\[CrossRef\]](#) [\[PubMed\]](#)

176. Colotta, F.; Re, F.; Polentarutti, N.; Suzzani, S.; Mantovani, A. Modulation of granulocyte survival and programmed cell death by cytokines and bacterial products. *Blood* **1992**, *80*, 2012–2020. [PubMed]
177. Mikirova, N.; Riondan, N.; Casciani, J. Modulation of Cytokines in Cancer Patients by Intravenous Ascorbate Therapy. *Mod. Sci. Monit.* **2016**, *22*, 14–25. [CrossRef] [PubMed]
178. Ferron-Celma, I.; Manilla, A.; Hassan, L.; Garcia-Navarro, A.; Comino, A.M.; Bueno, P.; Ferron, J.A. Effect of vitamin C administration on neutrophil apoptosis in septic patients after abdominal surgery. *J. Surg. Res.* **2009**, *153*, 224–230. [CrossRef] [PubMed]
179. Pechoux, R.D. With Friends like These: The Complex Role of Neutrophils in the Progression of Severe Pneumonia. *Front. Cell. Infect. Microbiol.* **2017**, *7*, 160. [CrossRef] [PubMed]
180. Zawrotniak, M.; Rapala-Kozik, M. Neutrophil extracellular traps (NETs)—Formation and implications. *Acta Biochim. Pol.* **2013**, *60*, 277–284. [PubMed]
181. Fuchs, T.A.; Abed, U.; Goosmann, C.; Haurwitz, R.; Schulze, I.; Wahn, V.; Weinrauch, Y.; Brinkmann, V.; Zychlinsky, A. Novel cell death program leads to neutrophil extracellular traps. *J. Cell Biol.* **2007**, *176*, 231–241. [CrossRef] [PubMed]
182. Brinkmann, V.; Reichard, U.; Goosmann, C.; Fauler, B.; Uhlemann, Y.; Weiss, D.S.; Weinrauch, Y.; Zychlinsky, A. Neutrophil extracellular traps kill bacteria. *Science* **2004**, *303*, 1532–1535. [CrossRef] [PubMed]
183. Parker, H.; Albert, A.M.; Kettle, A.J.; Winterbourn, C.C. Myeloperoxidase associated with neutrophil extracellular traps is active and mediates bacterial killing in the presence of hydrogen peroxide. *J. Leukoc. Biol.* **2012**, *91*, 369–376. [CrossRef] [PubMed]
184. Czaikoski, P.G.; Mota, J.M.; Nascimento, D.C.; Sonogo, F.; Castanheira, F.V.; Melo, P.H.; Scortegagna, G.T.; Silva, R.L.; Barroso-Sousa, R.; Souto, F.O.; et al. Neutrophil extracellular traps induce organ damage during experimental and clinical sepsis. *PLoS ONE* **2016**, *11*, e0148142. [CrossRef] [PubMed]
185. Camiciá, G.; Pozner, R.; de Larranaga, G. Neutrophil extracellular traps in sepsis. *Shock* **2014**, *42*, 286–294. [CrossRef] [PubMed]
186. Silk, E.; Zhao, H.; Weng, H.; Ma, D. The role of extracellular histone in organ injury. *Cell Death Dis.* **2017**, *8*, e2812. [CrossRef] [PubMed]
187. Margraf, S.; Loggers, T.; Reipen, J.; Altrichter, J.; Scholz, M.; Windolf, J. Neutrophil-derived circulating free DNA (cf-DNA/NETs): A potential prognostic marker for posttraumatic development of inflammatory second hit and sepsis. *Shock* **2008**, *30*, 352–358. [CrossRef] [PubMed]
188. Natarajan, R.; Fisher, B.J.; Syed, A.A.; Fowler, A.A. Impact of intravenous ascorbic acid infusion on novel biomarkers in patients with severe sepsis. *J. Palm. Respir. Med.* **2014**, *4*, 8. [CrossRef]
189. Elks, P.M.; van Eeden, F.J.; Dixon, G.; Wang, X.; Reyes-Aldasoro, C.C.; Ingham, P.W.; Whyte, M.K.; Walmsley, S.R.; Renshaw, S.A. Activation of hypoxia-inducible factor-1alpha (HIF-1alpha) delays inflammation resolution by reducing neutrophil apoptosis and reverse migration in a zebrafish inflammation model. *Blood* **2011**, *118*, 712–722. [CrossRef] [PubMed]
190. Hirota, K.; Semenza, G.L. Regulation of hypoxia-inducible factor 1 by prolyl and asparaginyl hydroxylases. *Biochem. Biophys. Res. Commun.* **2005**, *338*, 610–616. [CrossRef] [PubMed]
191. McInturff, A.M.; Cody, M.J.; Elliott, E.A.; Glenn, J.W.; Rowley, J.W.; Rondina, M.T.; Yost, C.C. Mammalian target of rapamycin regulates neutrophil extracellular trap formation via induction of hypoxia-inducible factor 1 alpha. *Blood* **2012**, *120*, 3118–3125. [CrossRef] [PubMed]
192. Hong, J.M.; Kim, J.H.; Kang, J.S.; Lee, W.J.; Hwang, Y.I. Vitamin C is taken up by human T cells via sodium-dependent vitamin C transporter 2 (SVCT2) and exerts inhibitory effects on the activation of these cells in vitro. *Anat. Cell Biol.* **2016**, *49*, 88–98. [CrossRef] [PubMed]
193. Bergsten, P.; Yu, R.; Kehrl, J.; Levine, M. Ascorbic acid transport and distribution in human B lymphocytes. *Arch. Biochem. Biophys.* **1995**, *317*, 208–214. [CrossRef] [PubMed]
194. Lenton, K.J.; Theriault, H.; Fulop, T.; Payette, H.; Wagner, J.R. Glutathione and ascorbate are negatively correlated with oxidative DNA damage in human lymphocytes. *Carcinogenesis* **1999**, *20*, 607–613. [CrossRef] [PubMed]
195. Campbell, I.D.; Cole, M.; Bunditratavorn, B.; Vella, A.T. Ascorbic acid is a potent inhibitor of various forms of T cell apoptosis. *Cell. Immunol.* **1999**, *194*, 1–5. [CrossRef] [PubMed]
196. Huijskens, M.J.; Walczak, M.; Sarkar, S.; Atrati, F.; Senden-Gijbbers, B.L.; Tinnus, M.G.; Bos, G.M.; Wieten, L.; Germeraad, W.T. Ascorbic acid promotes proliferation of natural killer cell populations in culture systems applicable for natural killer cell therapy. *Cytotherapy* **2015**, *17*, 613–620. [CrossRef] [PubMed]

197. Penn, N.D.; Purkins, L.; Kelleher, J.; Heatley, R.V.; Mascie-Taylor, B.H.; Belfield, P.W. The effect of dietary supplementation with vitamins A, C and E on cell-mediated immune function in elderly long-stay patients: A randomized controlled trial. *Age Ageing* **1991**, *20*, 169–174. [\[CrossRef\]](#) [\[PubMed\]](#)
198. Heuser, G.; Vojdani, A. Enhancement of natural killer cell activity and T and B cell function by buffered vitamin C in patients exposed to toxic chemicals: The role of protein kinase-C. *Immunopharmacol. Immunotoxicol.* **1997**, *19*, 291–312. [\[CrossRef\]](#) [\[PubMed\]](#)
199. Sasidharan Nair, V.; Song, M.H.; Oh, K.I. Vitamin C Facilitates Demethylation of the Foxp3 Enhancer in a Tet-Dependent Manner. *J. Immunol.* **2016**, *196*, 2119–2131. [\[CrossRef\]](#) [\[PubMed\]](#)
200. Nikolouli, E.; Hardtke-Wolenski, M.; Hapke, M.; Beckstette, M.; Geffers, R.; Floess, S.; Jaecel, E.; Huehn, J. Alkoxigen-Induced Regulatory T Cells Generated in Presence of Vitamin C Display Enhanced Stability of Foxp3 Expression and Promote Skin Allograft Acceptance. *Front. Immunol.* **2017**, *8*, 748. [\[CrossRef\]](#) [\[PubMed\]](#)
201. Morfort, A.; Wutz, A. Breathing-in epigenetic change with vitamin C. *EMBO Rep.* **2013**, *14*, 337–346. [\[CrossRef\]](#) [\[PubMed\]](#)
202. Song, C.X.; He, C. Potential functional roles of DNA demethylation intermediates. *Trends Biochem. Sci.* **2013**, *38*, 480–484. [\[CrossRef\]](#) [\[PubMed\]](#)
203. Johnston, C.S.; Barkyoumb, C.M.; Schumacher, S.S. Vitamin C supplementation slightly improves physical activity levels and reduces cold incidence in men with marginal vitamin C status: A randomized controlled trial. *Nutrients* **2014**, *6*, 2572–2583. [\[CrossRef\]](#) [\[PubMed\]](#)
204. Haryanto, B.; Sukmasari, T.; Wintergerst, E.; Maggini, S. Multivitamin supplementation supports immune function and ameliorates conditions triggered by reduced air quality. *Vitam. Miner.* **2013**, *4*, 1–15.
205. Romieu, I.; Castro-Giner, F.; Kunzli, N.; Sunyer, J. Air pollution, oxidative stress and dietary supplementation: A review. *Eur. Respir. J.* **2008**, *31*, 179–196. [\[CrossRef\]](#) [\[PubMed\]](#)
206. Kelly, F.; Dunster, C.; Mudway, I. Air pollution and the elderly: Oxidant/antioxidant issues worth consideration. *Eur. Respir. J.* **2003**, *21*, 706–756. [\[CrossRef\]](#)
207. Marmot, A.; Eley, J.; Stafford, M.; Starsfeld, S.; Warwick, E.; Marmot, M. Building health: An epidemiological study of “sick building syndrome” in the Whitehall II study. *Occup. Environ. Med.* **2006**, *63*, 283–289. [\[CrossRef\]](#) [\[PubMed\]](#)
208. Pozzer, A.; Zimmermann, P.; Doering, U.; van Aardenne, J.; Tost, H.; Dentener, F.; Janssens-Maenhout, G.; Lelieveld, J. Effects of business-as-usual anthropogenic emissions on air quality. *Atmos. Chem. Phys.* **2012**, *12*, 6915–6937. [\[CrossRef\]](#)
209. Sram, R.J.; Binkova, B.; Rossner, P., Jr. Vitamin C for DNA damage prevention. *Mutat. Res.* **2012**, *733*, 39–49. [\[CrossRef\]](#) [\[PubMed\]](#)
210. Tribble, D.; Giuliano, L.; Fortmann, S. Reduced plasma ascorbic acid concentrations in nonsmokers regularly exposed to environmental tobacco smoke. *Am. J. Clin. Nutr.* **1993**, *56*, 886–890. [\[PubMed\]](#)
211. Valkonen, M.; Kuusi, T. Passive smoking induces atherogenic changes in low-density lipoprotein. *Circulation* **1998**, *97*, 2012–2016. [\[CrossRef\]](#) [\[PubMed\]](#)
212. Schectman, G.; Byrd, J.C.; Hoffmann, R. Ascorbic acid requirements for smokers: Analysis of a population survey. *Am. J. Clin. Nutr.* **1991**, *53*, 1466–1470. [\[PubMed\]](#)
213. Preston, A.M.; Rodriguez, C.; Rivera, C.E.; Sahai, H. Influence of environmental tobacco smoke on vitamin C status in children. *Am. J. Clin. Nutr.* **2003**, *77*, 167–172. [\[PubMed\]](#)
214. Strauss, R. Environmental tobacco smoke and serum vitamin C levels in children. *Pediatrics* **2001**, *107*, 540–542. [\[CrossRef\]](#) [\[PubMed\]](#)
215. Panda, K.; Chattopadhyay, R.; Chattopadhyay, D.J.; Chatterjee, I.B. Vitamin C prevents cigarette smoke-induced oxidative damage in vivo. *Free Radic. Biol. Med.* **2000**, *29*, 115–124. [\[CrossRef\]](#)
216. Dietrich, M.; Block, G.; Benowitz, N.; Morrow, J.; Hudes, M.; Jacob, P., III; Norkus, E.; Packer, L. Vitamin C supplementation decreases oxidative stress biomarker 12-isoprostanes in plasma of nonsmokers exposed to environmental tobacco smoke. *Nutr. Cancer* **2003**, *45*, 176–184. [\[CrossRef\]](#) [\[PubMed\]](#)
217. Bagattkar, J.; Demuth, D.; Scott, D. Tobacco use increases susceptibility to bacterial infection. *Tob. Induc. Dis.* **2008**, *4*, 12. [\[CrossRef\]](#) [\[PubMed\]](#)
218. Ancavi, L.; Benowitz, N.L. Cigarette smoking and infection. *Arch. Intern. Med.* **2004**, *164*, 2206–2216. [\[CrossRef\]](#) [\[PubMed\]](#)

219. Sargeant, L.; Jeckel, A.; Wareham, N. Interaction of vitamin C with the relation between smoking and obstructive airways disease in EPIC Norfolk. European Prospective Investigation into Cancer and Nutrition. *Eur. Respir. J.* **2000**, *16*, 397–403. [\[CrossRef\]](#) [\[PubMed\]](#)
220. Peleg, A.Y.; Weeraratna, T.; McCarthy, J.S.; Davis, T.M. Common infections in diabetes: Pathogenesis, management and relationship to glycaemic control. *Diabetes Metab. Res. Rev.* **2007**, *23*, 3–13. [\[CrossRef\]](#) [\[PubMed\]](#)
221. Narayan, K.M.V.; Williams, D.; Gregg, E.W.; Cowie, C.C. (Eds.) *Diabetes Public Health: From Data to Policy*; Oxford University Press: Oxford, UK, 2011.
222. Pirota, L.; Ferraz, J. Role of pro- and anti-inflammatory phenomena in the pathophysiology of type 2 diabetes and obesity. *World J. Biol. Chem.* **2017**, *8*, 120–128. [\[CrossRef\]](#) [\[PubMed\]](#)
223. Donath, M. Targeting inflammation in the treatment of type 2 diabetes: Time to start. *Nat. Rev. Drug Discov.* **2014**, *13*, 465–476. [\[CrossRef\]](#) [\[PubMed\]](#)
224. Ferrante, A.W., Jr. Macrophages, fat, and the emergence of immunometabolism. *J. Clin. Investig.* **2013**, *123*, 4992–4993. [\[CrossRef\]](#) [\[PubMed\]](#)
225. Osborn, O.; Olefsky, J.M. The cellular and signaling networks linking the immune system and metabolism in disease. *Nat. Med.* **2012**, *18*, 363–374. [\[CrossRef\]](#) [\[PubMed\]](#)
226. Wilson, R.; Willis, J.; Geary, R.; Skidmore, P.; Fleming, E.; Frampton, C.; Carr, A. Inadequate vitamin C status in prediabetes and type 2 diabetes mellitus: Associations with glycaemic control, obesity, and smoking. *Nutrients* **2017**, *9*, 997. [\[CrossRef\]](#) [\[PubMed\]](#)
227. Maggini, S.; Wenzlaff, S.; Hornig, D. Essential role of vitamin C and zinc in child immunity and health. *J. Int. Med. Res.* **2010**, *38*, 386–414. [\[CrossRef\]](#) [\[PubMed\]](#)
228. Wintergerst, E.; Maggini, S.; Hornig, D. Immune-enhancing role of vitamin C and zinc and effect on clinical conditions. *Ann. Nutr. Metab.* **2006**, *50*, 85–94. [\[CrossRef\]](#) [\[PubMed\]](#)
229. Harding, A.H.; Wareham, N.J.; Bingham, S.A.; Khaw, K.; Luben, R.; Welch, A.; Forouhi, N.G. Plasma vitamin C level, fruit and vegetable consumption, and the risk of new-onset type 2 diabetes mellitus: The European prospective investigation of cancer—Norfolk prospective study. *Arch. Intern. Med.* **2008**, *168*, 1493–1499. [\[CrossRef\]](#) [\[PubMed\]](#)
230. Kositsawat, J.; Freeman, V.L. Vitamin C and A1c relationship in the National Health and Nutrition Examination Survey (NHANES) 2003–2006. *J. Am. Coll. Nutr.* **2011**, *30*, 477–483. [\[CrossRef\]](#) [\[PubMed\]](#)
231. Carter, P.; Gray, L.J.; Talbot, D.; Morris, D.H.; Khunti, K.; Davies, M.J. Fruit and vegetable intake and the association with glucose parameters: A cross-sectional analysis of the Let's Prevent Diabetes Study. *Eur. J. Clin. Nutr.* **2013**, *67*, 12–17. [\[CrossRef\]](#) [\[PubMed\]](#)
232. Mazloom, Z.; Hejazi, N.; Dabaghmanesh, M.H.; Tabatabaei, H.R.; Ahmadi, A.; Ansari, H. Effect of vitamin C supplementation on postprandial oxidative stress and lipid profile in type 2 diabetic patients. *Pak. J. Biol. Sci.* **2011**, *14*, 900–904. [\[PubMed\]](#)
233. Ashor, A.W.; Werner, A.D.; Lara, J.; Willis, N.D.; Mathers, J.C.; Siervo, M. Effects of vitamin C supplementation on glycaemic control: A systematic review and meta-analysis of randomised controlled trials. *Eur. J. Clin. Nutr.* **2017**. [\[CrossRef\]](#) [\[PubMed\]](#)
234. Hajishengallis, G. Too old to fight? Aging and its toll on innate immunity. *Mol. Oral Microbiol.* **2010**, *25*, 25–37. [\[CrossRef\]](#) [\[PubMed\]](#)
235. Cheng, L.; Cohen, M.; Bhugavan, H. Vitamin C and the elderly. In *CRC Handbook of Nutrition in the Aged*; Watson, R., Ed.; CRC Press Inc.: Boca Raton, FL, USA, 1985; pp. 157–185.
236. Simon, J.; Hudes, E.; Tice, J. Relation of serum ascorbic acid to mortality among US adults. *J. Am. Coll. Nutr.* **2001**, *20*, 255–263. [\[CrossRef\]](#) [\[PubMed\]](#)
237. Fletcher, A.; Breeze, E.; Shetty, P. Antioxidant vitamins and mortality in older persons: Findings from the nutrition add-on study to the Medical Research Council Trial of Assessment and Management of Older People in the Community. *Am. J. Clin. Nutr.* **2003**, *78*, 999–1010. [\[PubMed\]](#)
238. Thurman, J.; Mooradian, A. Vitamin supplementation therapy in the elderly. *Drugs Aging* **1997**, *11*, 433–449. [\[CrossRef\]](#) [\[PubMed\]](#)
239. Hanck, A. Vitamin C in the elderly. *Int. J. Vitam. Nutr. Res. Suppl.* **1983**, *24*, 257–269. [\[PubMed\]](#)
240. Schorah, C.J. The level of vitamin C reserves required in man: Towards a solution to the controversy. *Proc. Nutr. Soc.* **1981**, *40*, 147–154. [\[CrossRef\]](#) [\[PubMed\]](#)

241. Hunt, C.; Chakravorty, N.; Annan, G. The clinical and biochemical effects of vitamin C supplementation in short-stay hospitalized geriatric patients. *Int. J. Vitam. Nutr. Res.* **1984**, *54*, 65–74. [\[PubMed\]](#)
242. Mayland, C.R.; Bennett, M.L.; Allan, K. Vitamin C deficiency in cancer patients. *Palliat. Med.* **2005**, *19*, 17–20. [\[CrossRef\]](#) [\[PubMed\]](#)
243. Darai, P.A.; Moss, M.; Mannino, D.M.; Martin, G.S. The epidemiology of sepsis in patients with malignancy. *Chest* **2006**, *129*, 1432–1440. [\[CrossRef\]](#) [\[PubMed\]](#)
244. Gan, R.; Eintracht, S.; Hoffer, L.J. Vitamin C deficiency in a university teaching hospital. *J. Am. Coll. Nutr.* **2008**, *27*, 428–433. [\[CrossRef\]](#) [\[PubMed\]](#)
245. Bakaev, V.V.; Duntau, A.P. Ascorbic acid in blood serum of patients with pulmonary tuberculosis and pneumonia. *Int. J. Tuberc. Lung Dis.* **2004**, *8*, 263–266. [\[PubMed\]](#)
246. Hunt, C.; Chakravorty, N.K.; Annan, G.; Habibzadeh, N.; Schorah, C.J. The clinical effects of vitamin C supplementation in elderly hospitalised patients with acute respiratory infections. *Int. J. Vitam. Nutr. Res.* **1994**, *64*, 212–219. [\[PubMed\]](#)
247. Bharara, A.; Grossman, C.; Grinnan, D.; Syed, A.A.; Fisher, B.J.; DeWilde, C.; Natarajan, R.; Fowler, A.A. Intravenous vitamin C administered as adjunctive therapy for recurrent acute respiratory distress syndrome. *Crit. Care* **2016**, *2016*, 8560871. [\[CrossRef\]](#) [\[PubMed\]](#)
248. Fowler, A.A.; Kim, C.; Lepler, L.; Malhotra, R.; DeBesa, O.; Natarajan, R.; Fisher, B.J.; Syed, A.; DeWilde, C.; Priday, A.; et al. Intravenous vitamin C as adjunctive therapy for enterovirus/rhinovirus induced acute respiratory distress syndrome. *World J. Crit. Care Med.* **2017**, *6*, 85–90. [\[CrossRef\]](#) [\[PubMed\]](#)
249. Hemila, H.; Chalker, E. Vitamin C for preventing and treating the common cold. *Cochrane Database Syst. Rev.* **2013**, *1*, CD00980. [\[PubMed\]](#)
250. Hemila, H. Vitamin C and the common cold. *Br. J. Nutr.* **1992**, *67*, 3–16. [\[CrossRef\]](#) [\[PubMed\]](#)
251. Hume, R.; Weyers, E. Changes in leucocyte ascorbic acid during the common cold. *Scott. Med. J.* **1973**, *18*, 3–7. [\[CrossRef\]](#) [\[PubMed\]](#)
252. Wilson, C.W. Ascorbic acid function and metabolism during colds. *Ann. N. Y. Acad. Sci.* **1975**, *258*, 529–539. [\[CrossRef\]](#) [\[PubMed\]](#)
253. Schwartz, A.R.; Togo, Y.; Hornick, R.B.; Tominaga, S.; Gleckman, R.A. Evaluation of the efficacy of ascorbic acid in prophylaxis of induced rhinovirus 44 infection in man. *J. Infect. Dis.* **1973**, *128*, 500–505. [\[CrossRef\]](#) [\[PubMed\]](#)
254. Davies, J.E.; Hughes, R.E.; Jones, E.; Reed, S.E.; Craig, J.W.; Tyrrell, D.A. Metabolism of ascorbic acid (vitamin C) in subjects infected with common cold viruses. *Biochem. Med.* **1979**, *21*, 78–85. [\[CrossRef\]](#)
255. Mochalkin, N. Ascorbic acid in the complex therapy of acute pneumonia. *Voen. Med. Zhurnal* **1970**, *9*, 17–21. Available online: <http://www.mvhselinski.fi/home/hemila/TS.pdf> (accessed on 5 December 2014).
256. Hemila, H.; Louhiala, P. Vitamin C for preventing and treating pneumonia. *Cochrane Database Syst. Rev.* **2013**, *5*, CD005332. [\[PubMed\]](#)



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